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What Open–Lung biopsy teaches us about ARDS in severe COVID–19 patients: Mechanisms, pathology and therapeutic implications.

THESIS

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HIPPOCRATIC OATH

AS A MEMBER OF THE MEDICAL PROFESSION:

*I SOLEMNLY PLEDGE to dedicate my life to the service of
humanity;*

*THE HEALTH AND WELL-BEING OF MY PATIENT will be
my first consideration;*

I WILL RESPECT the autonomy and dignity of my patient;

I WILL MAINTAIN the utmost respect for human life;

*I WILL NOT PERMIT considerations of age, disease or
disability, creed, ethnic origin, gender, nationality, political
affiliation, race, sexual orientation, social standing, or any
other factor to intervene between my duty and my patient;*

*I WILL RESPECT the secrets that are confided in me, even
after the patient has died;*

*I WILL PRACTISE my profession with conscience and dignity
and in accordance with good medical practice;*

*I WILL FOSTER the honour and noble traditions of the
medical profession;*

*I WILL GIVE to my teachers, colleagues, and students the
respect and gratitude that is their due;*

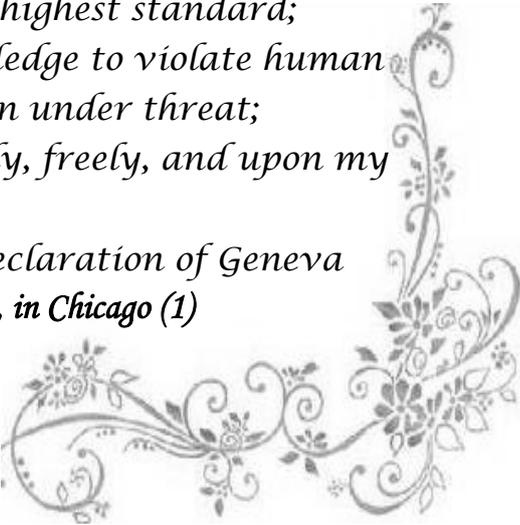
*I WILL SHARE my medical knowledge for the benefit of the
patient and the advancement of healthcare;*

*I WILL ATTEND TO my own health, well-being, and abilities
in order to provide care of the highest standard;*

*I WILL NOT USE my medical knowledge to violate human
rights and civil liberties, even under threat;*

*I MAKE THESE PROMISES solemnly, freely, and upon my
honour.*

*A newly revised version of the Declaration of Geneva
was adopted on October 14, 2017, in Chicago (1)*





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LISTE ARRÊTÉE LE 23/06/2021



DEDICATIONS

*“Nothing in life is to be feared, it is only to be understood.
Now is the time to understand more, so that we may fear less.”*

— Marie Curie —



I dedicate this thesis to...

To the memory of my grand-parents:

Malika Hadhoumi, Halima Koundi, Mohammed Abourida.

I wish I could go back in time and spend more time with you. I hope I could honour you today by dedicating this work to you. You were my role models and my source of benediction and blessing.

May your souls rest in peace.

To my hero baba Abdelaziz Abourida,

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Four ornate, black-and-white decorative corner ornaments are arranged in a square pattern around the central text. Each ornament features intricate scrollwork and floral motifs, with a small wavy line extending from the outer corner.

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To Professor Mohammed-Abdnasser Samkaoui,

Chairman of my thesis

Head of Anaesthesiology & Intensive Care Department

I am delighted to be granted this great honour by accepting the presidency of this committee. I thank you for accepting me as a volunteer in your department doing research work, and for giving me this remarkable unprecedented chance. I have always admired your human qualities and your professional skills, as I highly look up to you. Please accept, through this work, the expression of my gratitude and my deepest respect.

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Supervisor of my thesis

Head of Pathology Department

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To Professor Houssam Rebaï,

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To Professor Hichame Fenane,

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ABBREVIATIONS

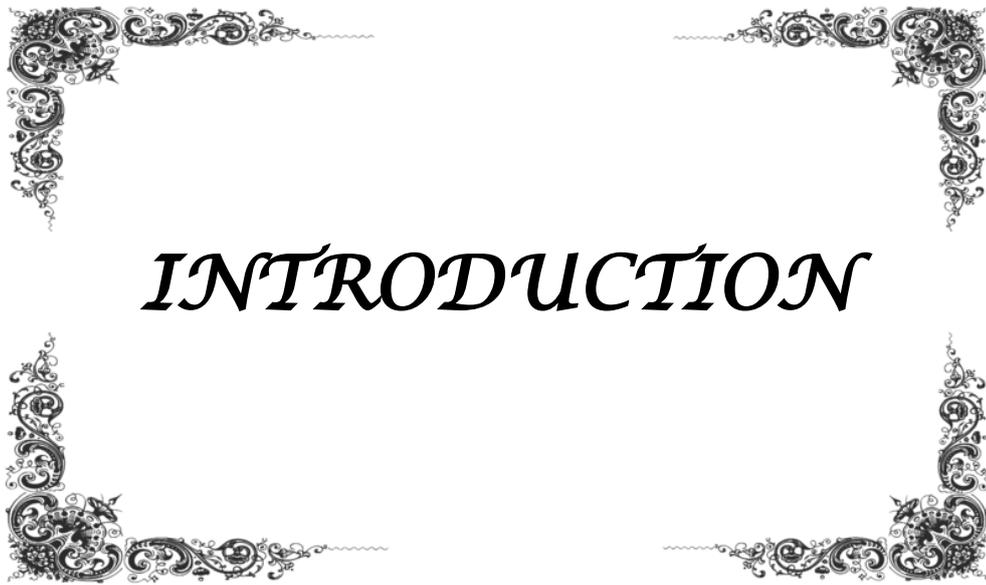
List of abbreviations

SARS-CoV-2:	Severe Respiratory Coronavirus 2
COVID-19	: Coronavirus Disease 2019
ICU	: Intensive Care
ARDS	: Acute Respiratory Distress Syndrome
DAD	: Diffuse alveolar damage
CT	: Computed tomography
PPE	: Personal Protective Equipment
RR	: Respiratory Rate
HR	: Heart Rate
SpO2	: Pulsed Oxygenation Saturation
Bp	: Blood Pressure
CPAP	: Continuous Positive Airway Pressure
IV	: Invasive Ventilation
µg	: Microgram
ML	: Millilitre
pg	: Picogramm
Mg	: Milligram
IL	: Interleukin
RT-PCR	: Reverse Transcription Polymerase Chain Reaction
PEEP	: Positive End-Expiratory pressure
ALI	: Acute Lung Injury
PaO2	: Partial pressure of oxygen
FiO2	: Fraction of inspired oxygen
MmHg	: Millimetre of mercury
AECC	: American-European Consensus Conference
MERS	: Middle East Respiratory Syndrome
ACE 2	: Angiotensin-converting enzyme 2
AT1-R	: Angiotensin II type 1 receptor
TNF	: Tumour Necrosis Factor
VWF	: Von Willebrand factor
CRP	: C-reactive protein
LMWH	: Low-molecular-weight heparin
RNA	: Ribonucleic Acid



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INTRODUCTION

In the early months of 2020, the world witnessed an outbreak of the Severe Respiratory Syndrome Coronavirus (SARS-CoV-2), which caused a tremendous flood of coronavirus-related pneumonia. In most cases, coronavirus disease 2019 (COVID-19) was rapidly resolved, whereas 26% require intensive care unit admission (ICU) (2). Clinical manifestations of COVID-19 vary from mild pneumonia to progressive Acute Respiratory Distress Syndrome (ARDS). Its singular features are severe hypoxemia often associated with near-normal respiratory system compliance at the beginning (3). On these distinctive grounds, a crucial question rose: How does COVID-19 damage the lungs to cause a rapidly progressive onset of profound hypoxemia?

In 1967, Ashbaugh et al. assumed that Diffuse Alveolar Damage (DAD) on lung histology was the pathological hallmark of ARDS (4). However, recent shreds of evidence point out that DAD is only a phenotype of ARDS among others, but with higher mortality (5). COVID-19 is a systemic disease that affects multiple organs, including the lungs, pharynx, heart, liver, brain, and kidneys (6). Very little is known about the “weaponry” of COVID-19; however, its main target seems to be the vascular endothelium. Initial reports documented clinically significant coagulopathy in critically ill patients (7) (8). According to the Berlin definition (9), diagnosing ARDS does not take into account pathologic findings which leaves a considerable gap in categorizing COVID-19 related ARDS (C-ARDS) and its peripheral vascular changes. On the other hand, previous work studied the co-infection rate between SARS-CoV-2 and other respiratory pathogens and focused only on nasopharyngeal swabs (10) but failed to address it in the lower respiratory tract.

Understanding the precise pathophysiology of C-ARDS will assist researchers and physicians in tailoring a timely-appropriated therapeutic approach. Here, we conducted a descriptive study performing an open-lung biopsy (OLB) in invasively ventilated patients with C-ARDS; taking into account the benefit-to-risk ratio. This study is aimed at determining C-ARDS pathological characteristics and co-infection with other pathogens in lung tissue.

Four decorative corner ornaments, each featuring intricate floral and scrollwork patterns, positioned at the corners of the page to frame the central text.

*MATERIALS
AND METHODS*

I. Patients and diagnosis

We selected patients with laboratory-confirmed SARS-CoV-2 infection –admitted to the COVID-19 ICU of the Moammed VI University Hospital of Marrakech, from the 25th of April to 25th June 2020– who later developed ARDS that met the Berlin definition (9) and were put under mechanical ventilation. Initial chest Computed Tomography (CT) scans revealed bilateral diffused ground-glass opacities in different percentages. Informed written consent from the next of kin was obtained. The ethics committee of the University Hospital approved this research respecting the regulations of the Helsinki declaration.

II. Surgical Technique

An open lung mini-thoracotomy with rib spreading was performed using the wedge resection technique from the anterolateral segment with a stapler (*Fig 1*). The anterior end of the incision was placed 3 to 4 cm lateral to the middle line of the breastbone. Pleural space was entered above the fifth rib. A chest tube was placed through another incision and the muscle layers were loosely closed with a running absorbable number 0 suture. A lung tissue fragment was immediately soaked in 4% formalin solution and the other fragments along with pleural effusion fluid were put in a culture environment. The surgical team wore level 3 personal protective equipment (PPE) during the invasive procedure (*Fig 2*)



Figure 1: Rib spreading during mini-thoracotomy.



Figure 2: Surgical team performing open-lung biopsy while wearing appropriate PPE.

III. Specimen's analysis

Rigorous steps were respected to assure biosecurity of pathological tissue samples and to minimize staff contamination, as portrayed (*Fig. 3*). Biopsy lung tissue was analyzed with hematoxylin-eosin, Periodic Acid Schiff for detecting bacterial and fungal infection, and also Masson Trichrome staining to identify pulmonary interstitial fibrosis. The slides have been digitized using Leica SCN400 Slide Scanner, and then images of tissue sections were captured. Pleural effusion fluid and biopsy lung tissue were tested for SARS-CoV-2 by RT-PCR and a panel of non-SARS-CoV-2 respiratory viral pathogens (Adenovirus, Coronavirus Metapneumovirus, Enterovirus, Rhinovirus, MERS-CoV, Parainfluenza virus, Syncytial respiratory virus, Flu virus A & B), along with standard bacterial and fungal respiratory cultures as shown (*Fig 4*).



Figure 4: Specimen collection materials (A: Pathology vial, B: Virology PCR vial, C: Sars-CoV-2 PCR vial, D: Bacteriology vial).



Figure 3: Biosecurity of pathological tissue samples (A: Photographing the pathological examination request form; B: Placing the vials containing the samples in a container; C: Spraying the bottle with a disinfectant solution; D: Packaging in a level II biological safety cabinet without removing the sample from the vial; E: Leaving to set for at least 24 hours in 4% formalin solution; F: Using a microbiological safety station and PPE.)

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RESULTS

I.

Clinical features

An open lung biopsy in 3 patients with C-ARDS was carried out. All of them were male and tested positive for SARS-CoV-2 by nasopharyngeal swab at the time of hospital admission. The median age was 65 years (range, 57-72 years). The median duration from symptoms to admission is 10 days (range, 7-13days), and the median duration from admission to death was 9.6 days (range, 5-15 days). Initial symptoms in 3 patients were mainly fever, dry cough, and shortness of breath, whereas Case 3 reported anosmia.

Hypertension was found in all patients as pre-existing comorbidity, Case 1 had hyperthyroidism and benign hypertrophy of the prostate, and Case 3 reported being a chronic smoker for 20 years with dyslipidemia. (*Table I*).

All patients were managed with the same national Moroccan protocol, which is hydroxychloroquine combined with azithromycin, Zinc, Vitamin C, and a therapeutic dose of Low Molecular Weight Heparin (LMWH), in addition to acetylsalicylic acid. Two patients had A⁺ blood type and Case 1 had O⁺. The median duration of non-invasive ventilation management was 6.3 days (range, 3-10 days) and the median duration of mechanical ventilation was 3.3 days (range, 2-5 days). Open lung biopsy was performed on the first day of endotracheal intubation in all patients.

Table I. Clinical, radiological and microbiological characteristics of patients, and treatment received during ICU stay.

	Case 1	Case 2	Case 3
Age,years	72	68	59
Gender	Male	Male	Male
Known comorbidities	Hypertension Hyperthyroidism Benign hypertrophy of the prostate	Hypertension	Hypertension Chronic Smoker Dyslipidemia
Symptoms	Fever 38,7 °C Dry cough Shortness of breath Thoracic pain	Fever 38,9 °C Dry cough Shortness of breath Fatigue	Fever 38,5°C Dry cough Anosmia
Symptom duration before admission, (days)	13	7	10
Admission to death, (days)	15	5	9
Physical examination in admission	RR=33cpm SpO ₂ =73% HR = 95 bpm BP: 110/60 mmHg Blood sugar: 1.44g/L	RR=36cpm SpO ₂ =84% HR=95cpm BP=120/60 mmHg Blood sugar: 1.7g/L	RR=34cpm SpO ₂ =83% HR=88cpm BP=130/80 mmHg Blood sugar: 3.5 g/L
THORACIC CT SCAN	Ground glass Crazy paving >70%	Ground glass >75%	Ground glass >80%
Blood culture	Multiresistant Acinetobacter Baumani Gram-Positive Bacteria sensitive to Colistin (Catheter-related infection)	Sterile	Sterile
Duration of ventilator management,(days)	CPAP = 10 IV= 5	CPAP = 3 IV=2	CPAP = 6 IV = 3
Treatment	Hydroxychloroquine 200mg x 3/24h Antibiotics (Azithromycin 250mg/24h + Ceftriaxone 2g/24h + Moxifloxacin 400mg/12h) Therapeutic dose of LMWH (enoxaparin 1 mg/kg x 2 /24h) Acetylsalicylic acid 160mg/24h Methylprednisolone 120mgx2/24h Zinc 220mg /24h Vitamin C 1,5g /24h Acetaminophen 500mg /6h		
	Carbimazole 10mg/24h Nebulised Colistin 2Million Unit x3/24h Tocilizumab 8 mg/kg ONCE		

Table I. Clinical, radiological and microbiological characteristics of patients, and treatment received during ICU stay." suite"

	Case 1	Case 2	Case 3
Blood type	O+	A+	A+
Microbiology (lung tissue and pleural fluid)	NEGATIVE	NEGATIVE	NEGATIVE
SARS-CoV-2 on Pleural fluid	Negative	Positive	Positive
SARS-CoV-2 on lung biopsy	Negative	Positive	Positive

RR: Respiratory rate, HR: Heart Rate SpO₂: Pulsed Oxygenation Saturation, BP: Blood Pressure, CPAP: Continuous Positive Airway Pressure, IV: Invasive Ventilation

II. Biological results

D-dimer serum levels at the admission of case 1,2, and 3 were 7.41 µg/mL, 2.27 µg/mL and 0.31 µg/mL respectively, whereas at their last day were elevated to 21.27 µg/mL , 22.95 µg/mL and 5 µg/mL (*Fig. 5*) . It's difficult to claim that none of the patients had thromboembolic events as no autopsy was performed.

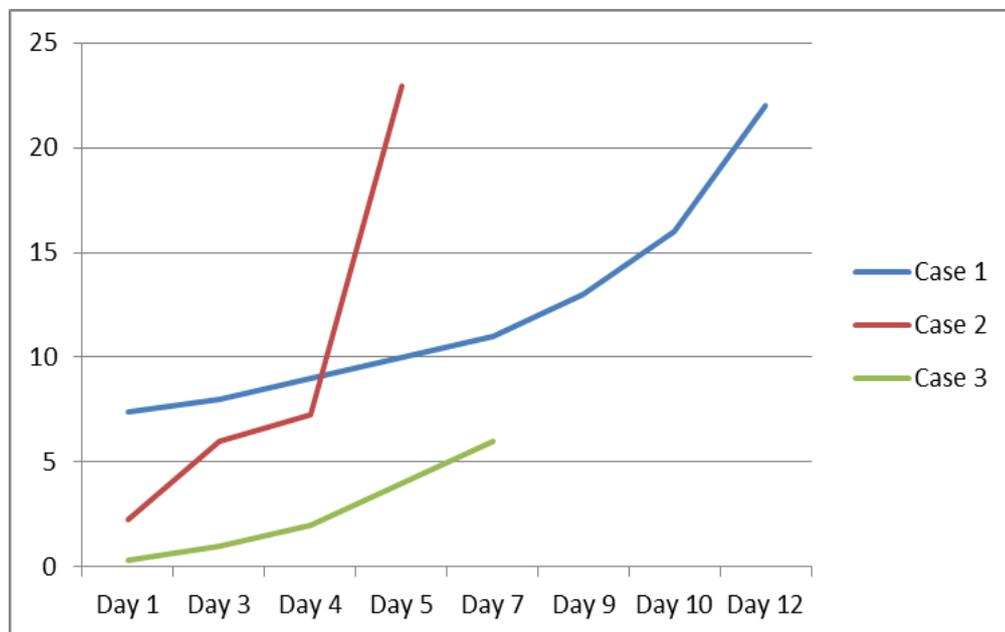


Figure 5: D-dimer level in patient's serum during ICU stay (µg/mL).

IL-6 serum levels at the admission of case 1,2, and 3 were 201 pg/mL, 107 μ g/mL and 31 pg/mL respectively, whereas prior to their last day were elevated to 675 pg/mL , 325 pg/mL and 280 pg/mL (Fig. 6).

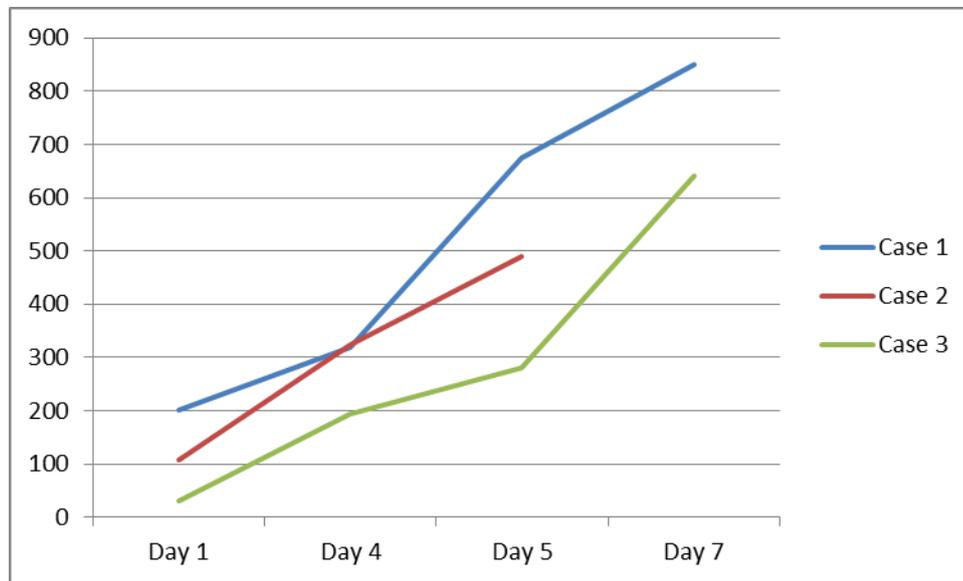


Figure 6: IL-6 level in patient's serum during ICU stay (pg/mL).

III. Microbiological results

Bacterial and viral (other than SARS-CoV-2) culture returned negative. Qualitative RT-PCR detected SARS-CoV-2 in the pulmonary parenchyma and pleural fluid of Case 2 and 3.

IV.

Histological results

Histological examination showed diffuse alveolar damage with collapsed alveoli (Fig. 8) and intensified thickening of the intercellular septa in Case 1 and 2, whereas Case 3 exhibited enlarged airspaces (Fig. 7), consistent with emphysema. The lumen was filled with proteinaceous and fibrin exudates (Fig. 12). Type II pneumocytes were found hyperplastic with an atypical appearance, multinucleated with enlarged and prominent nuclei (Fig. 10). There were significant focal points of pneumocyte desquamation, multinucleated giant cells, and hyaline membrane formation on the alveolar wall (Fig. 11). The interstitial tissue displayed oedema (Fig. 9) and widespread inflammatory infiltrates (Fig. 15) marked with lymphocytes mainly but also plasma cells, macrophages, and eosinophilic polynuclear cells. Prominent **microthrombi** (Fig. 16) and vascular congestion (Fig. 17) were the major pathological finding in all cases. Also, fibrin deposits (Fig. 14) were found in the vessel intima with a thickened vessel wall. Anthracosis deposit was also seen in Cases 1 and 2. No malignant tumour proliferation and no alveolar fibrosis (Fig. 18) were found in the 3 cases.

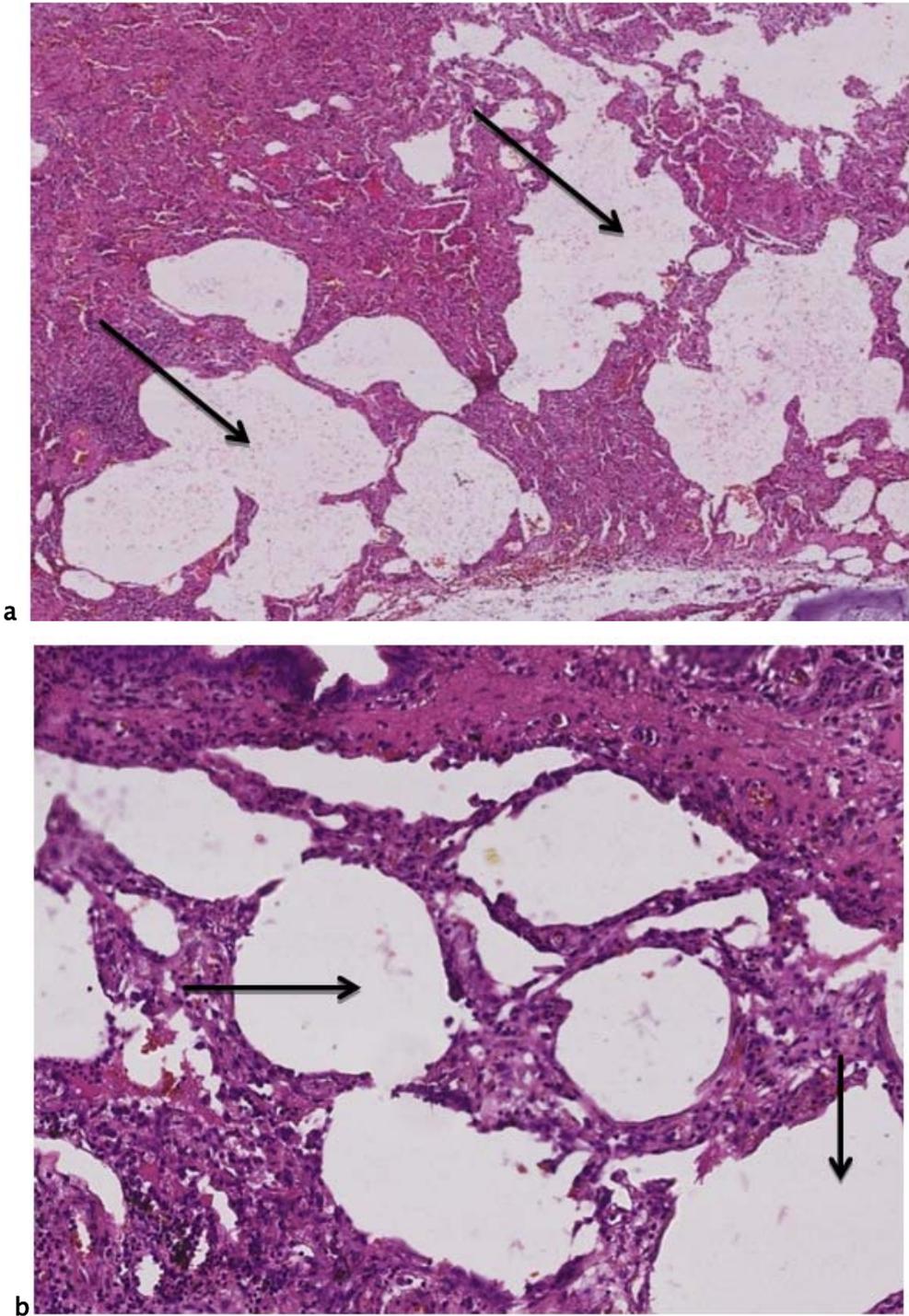


Figure 7: Alveolar dilatation (×20 magnification)

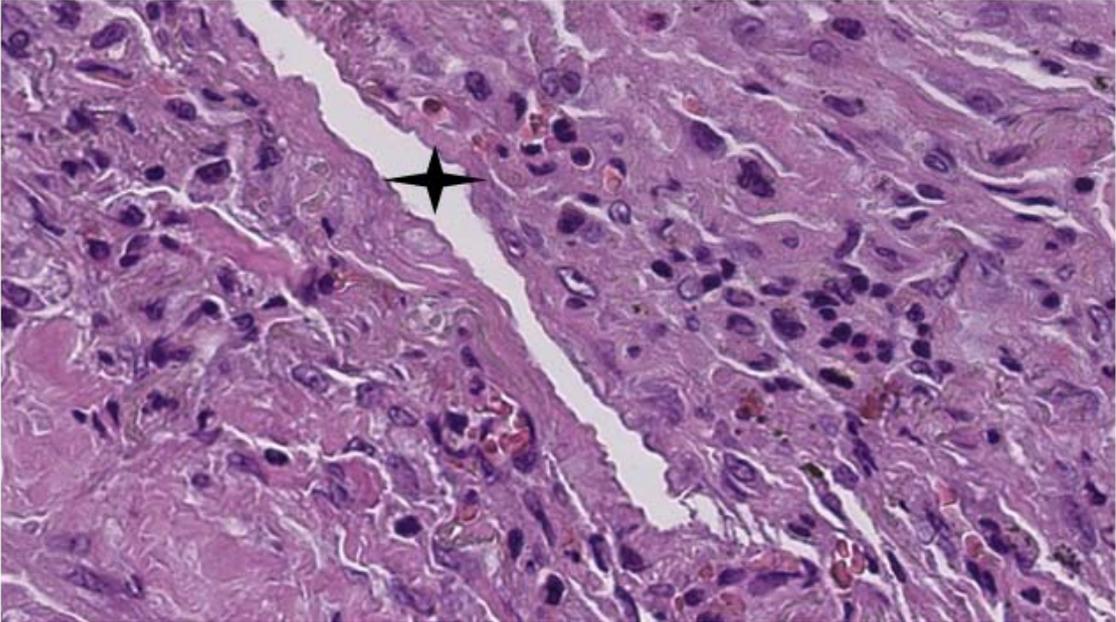


Figure 8: Collapsed alveoli. ($\times 40$ magnification).

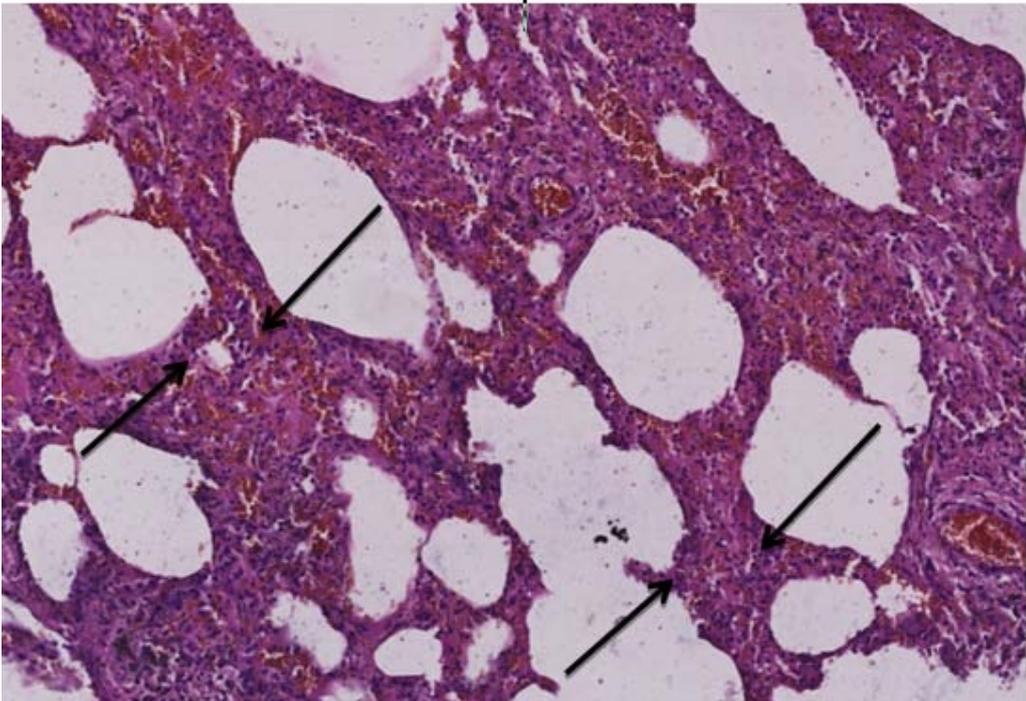


Figure 9: Inter-alveolar septum thickening ($\times 20$ magnification).

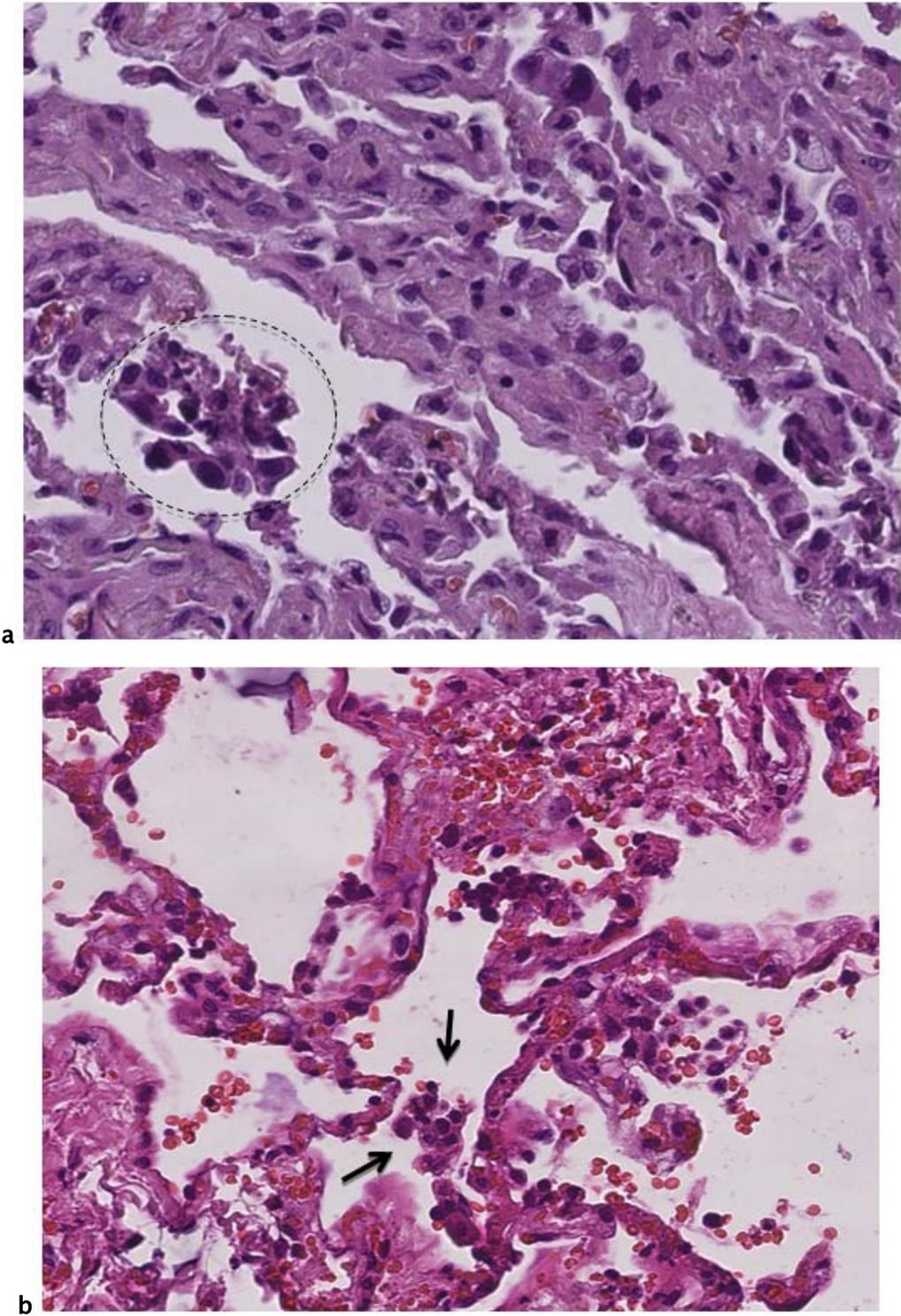
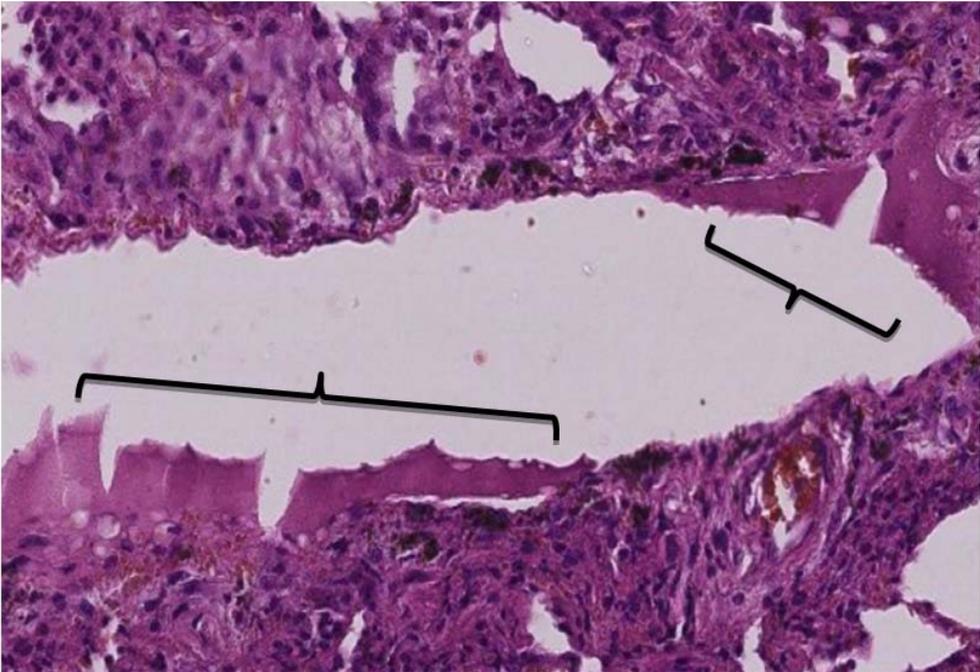
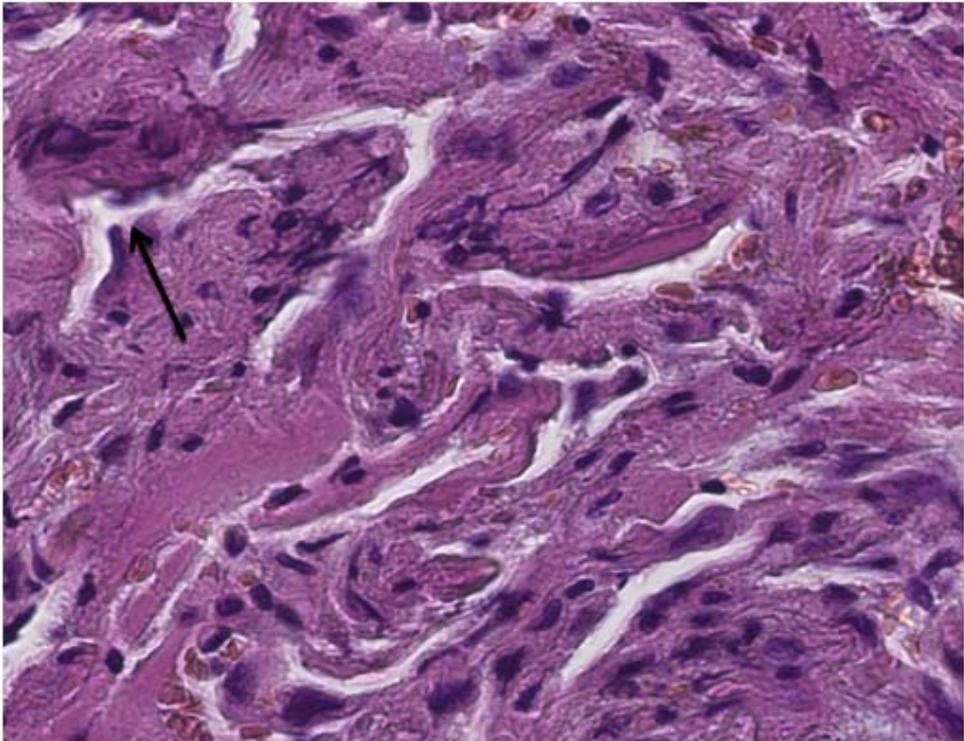


Figure 10: Epithelial desquamation and Type II pneumocyte hyperplasia (a,b x30 magnifications).



a



b

Figure 11: Hyaline membranes made of dead pneumocytes, surfactant and protein exudates. (a: x30magnification, b : x40 magnification)

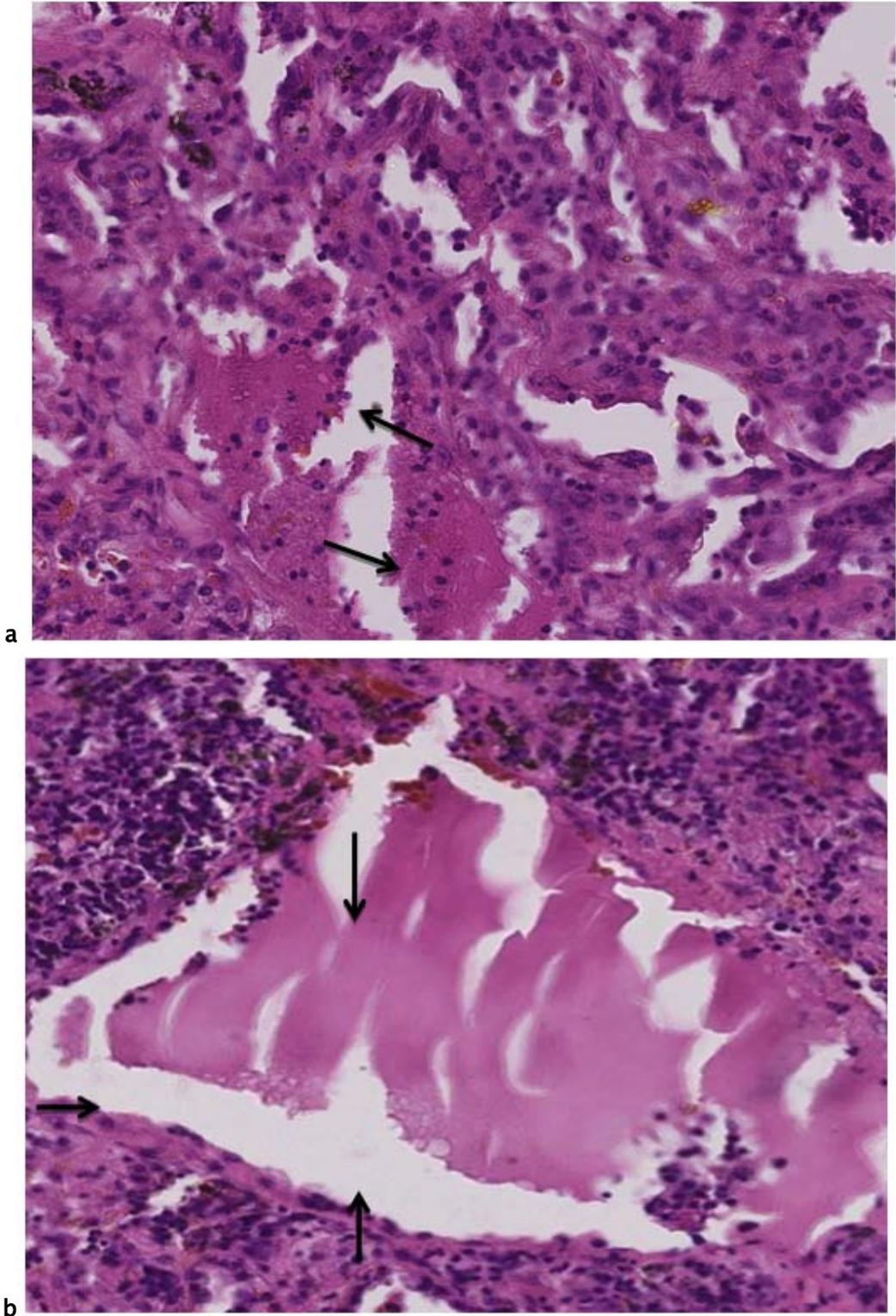


Figure 12: Exudate filling the alveolar cavity (a: x10magnification; b: x40magnification)

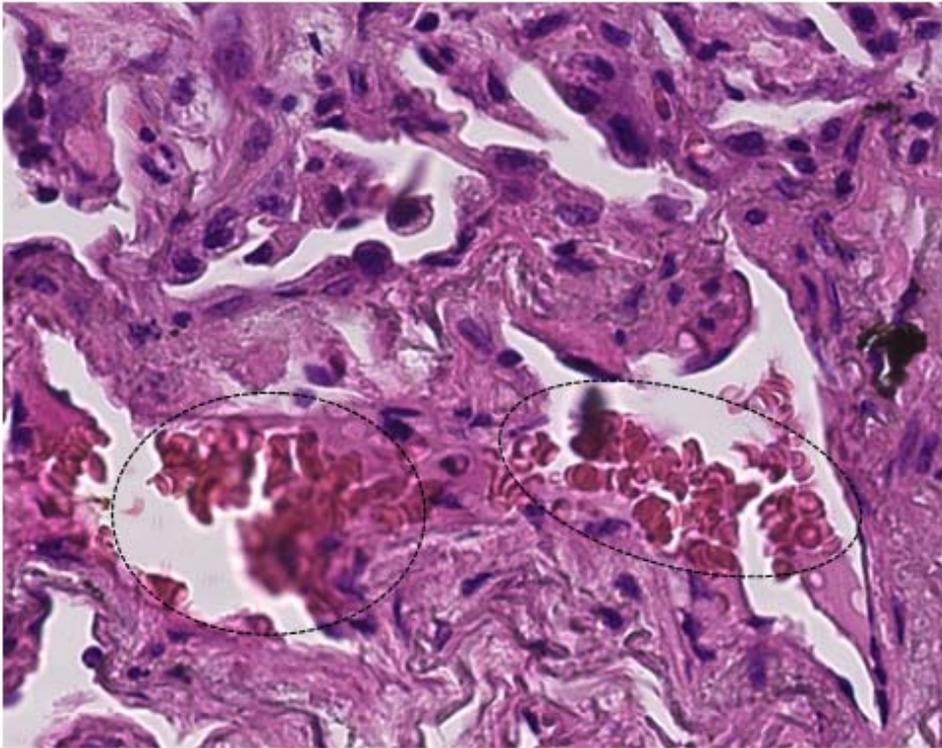


Figure 13: Alveolar haemorrhage (x20 magnification)

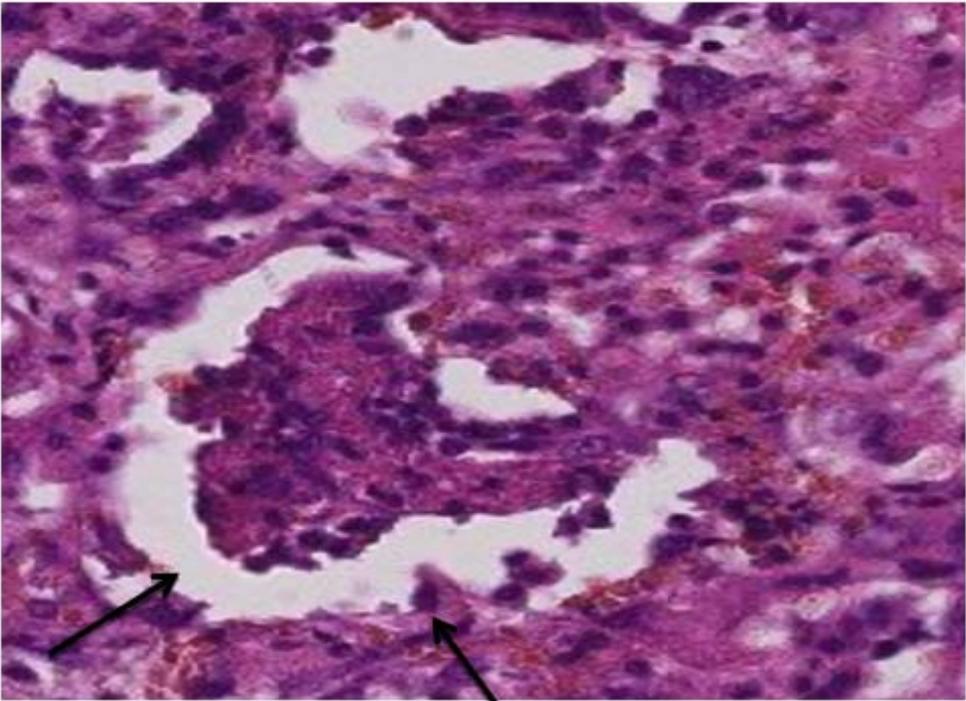


Figure 14: Intra-alveolar fibrin deposit (x40 magnification)

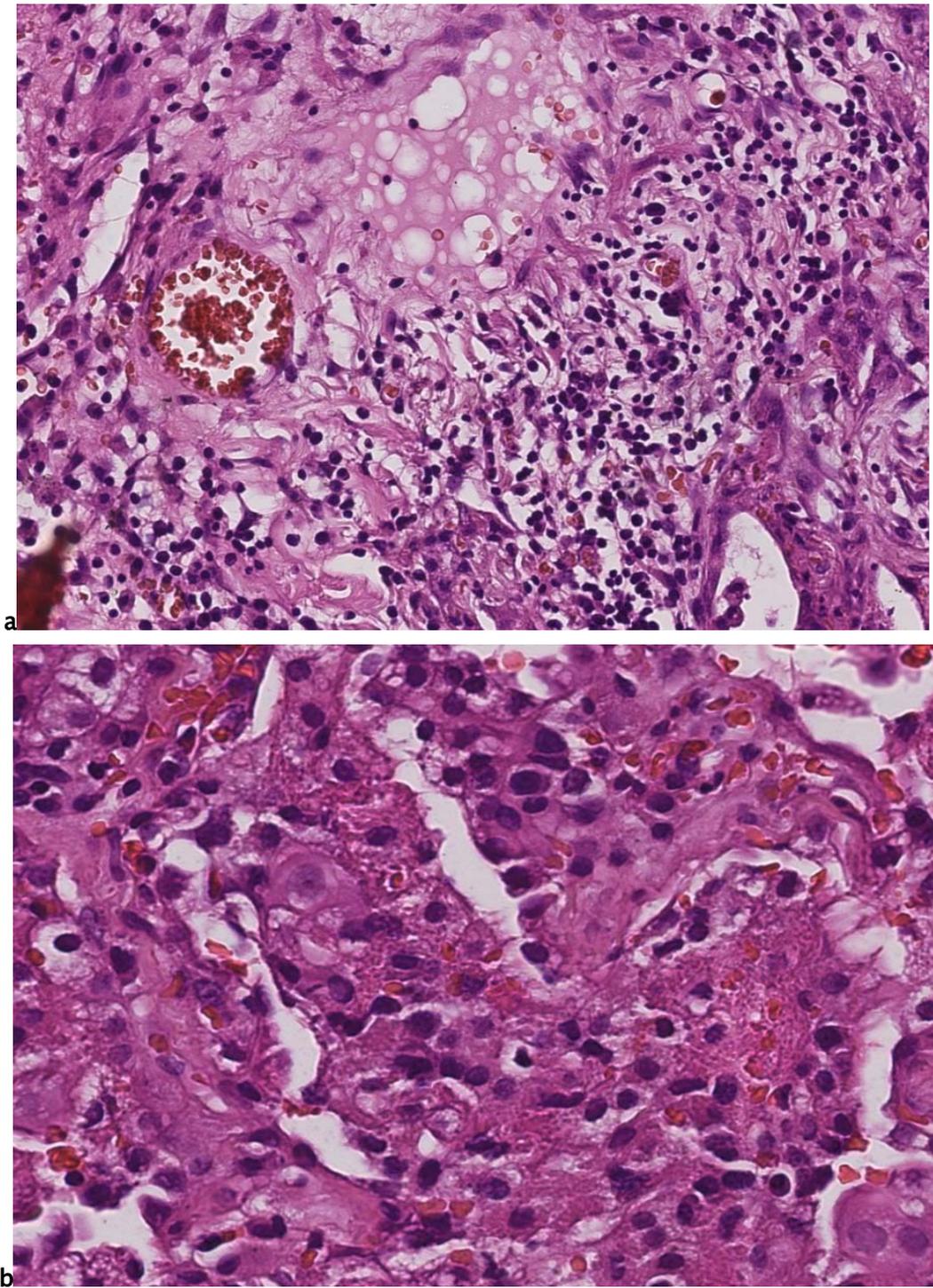


Figure 15: Intense polymorphic inflammation (a: x20 magnification, b: x40 magnification)

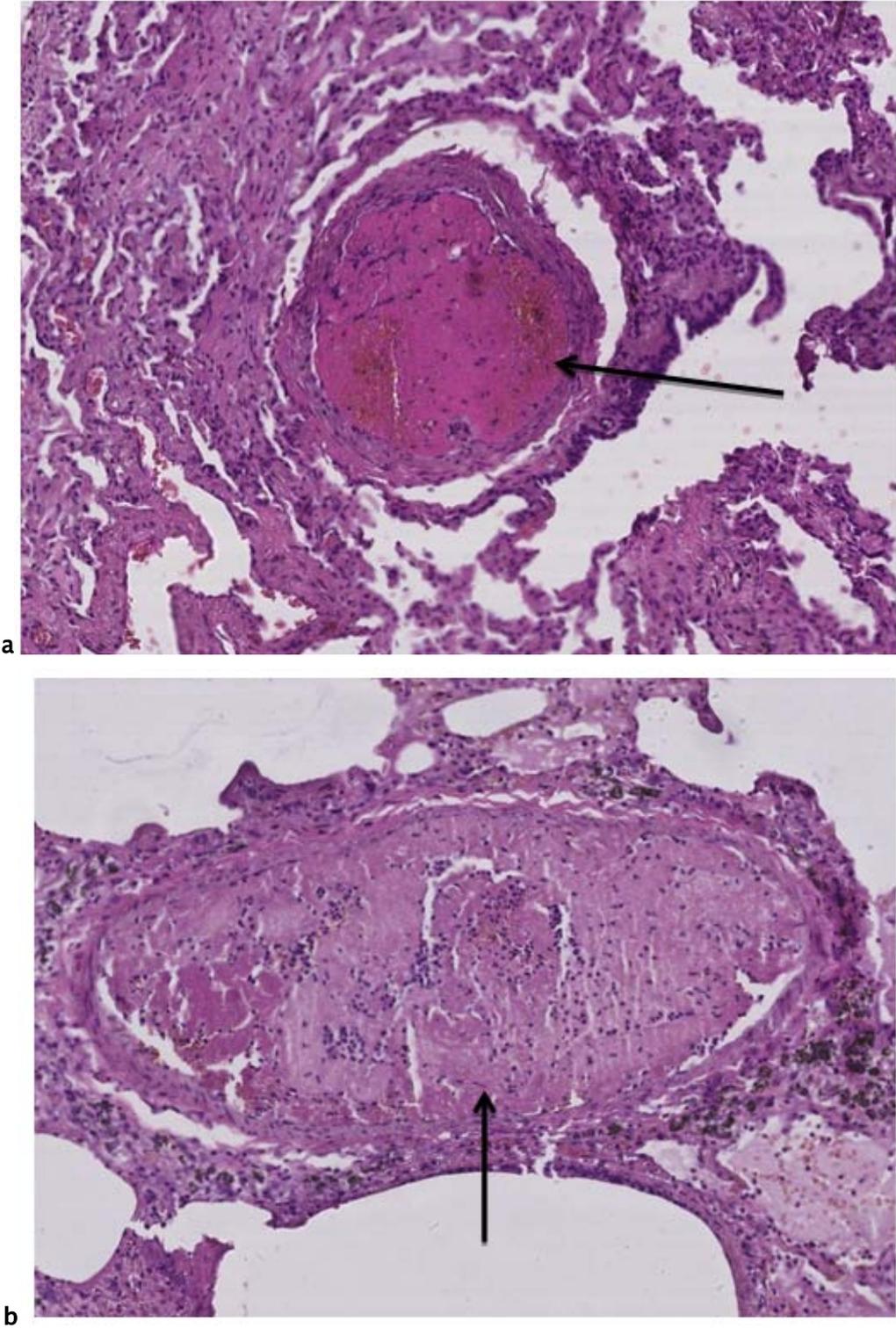


Figure 16: Prominent microthrombi (a, b : x20 magnification)

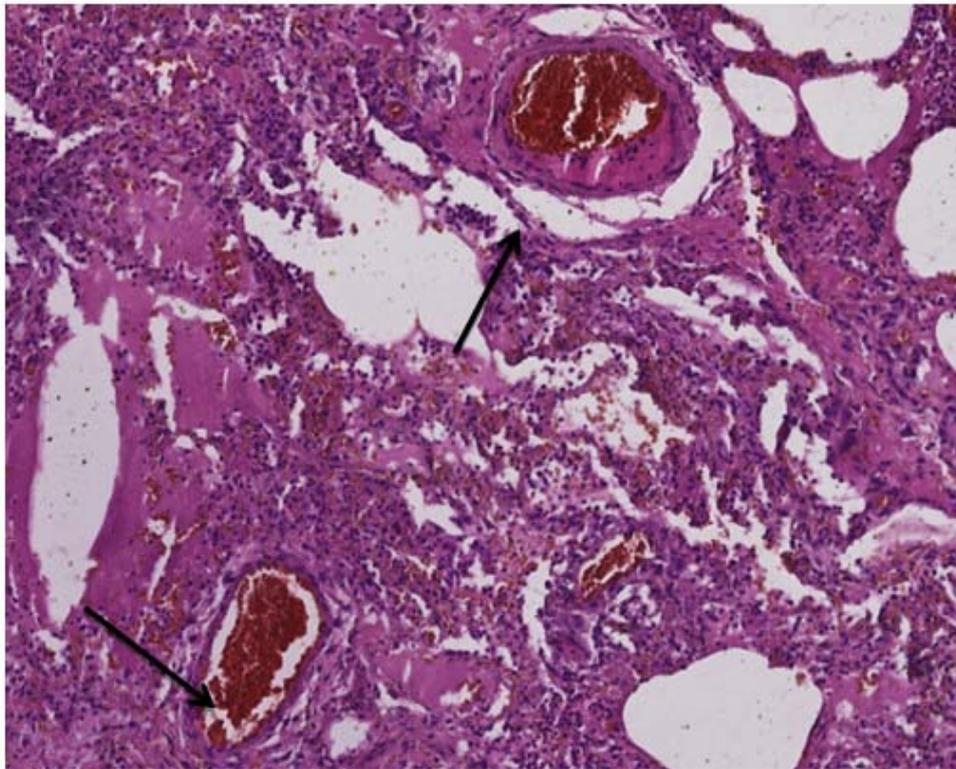


Figure 17: Vascular congestion (x 20 magnification)

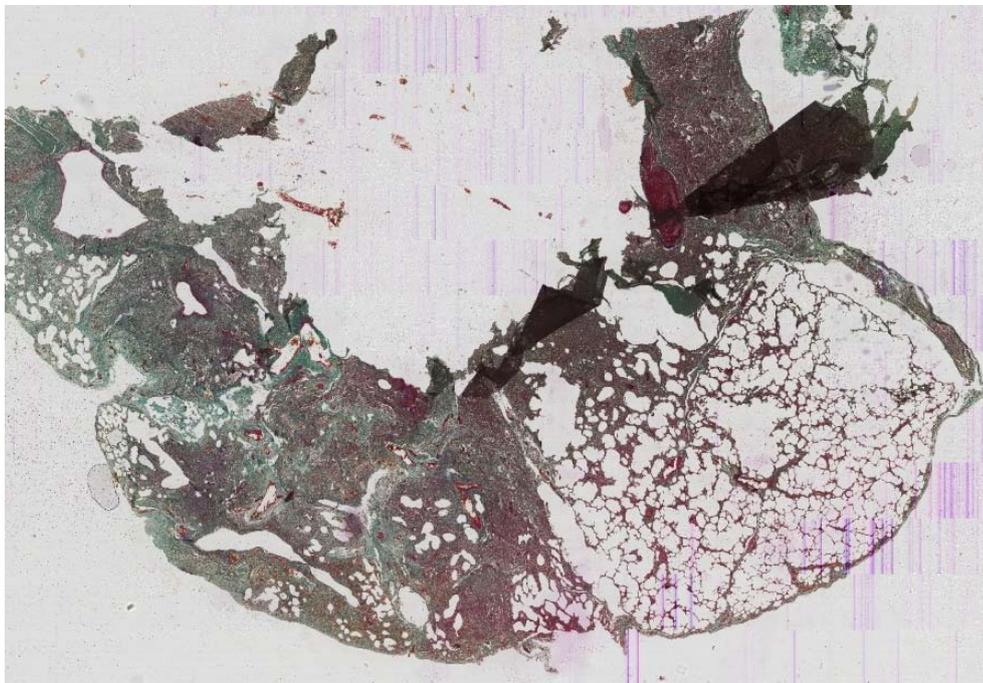


Figure 18: Trichrome Masson Coloration showing no pulmonary fibrosis.

Table II : Histological features of lung biopsy patients.

	Case 1	Case 2	Case 3
Alveoli	Variable size Collapsed +++	Variable size Collapsed +++ Enlarged +	Enlarged +++ Collapsed +
Inter-alveoli wall	Thickened +++	Thickened +++	Thickened ++ Dystrophic +
Type II pneumocyte	➤ Hyperplasic +++ ➤ Atypical ➤ Multinucleated ➤ Enlarged	➤ Hyperplasic +++ ➤ Atypical ➤ Multinucleated ➤ Enlarged	➤ Discontinuous ➤ Hyperplasic ++
Alveolar Cavity : • Hyaline Membrane • Exudate • Alveolar haemorrhage	+++ +++ ++	+++ +++ 0	++ ++ +
Interstitial Tissue : • Inflammatory infiltrate	➤ Diffused ➤ Minimal ➤ Lymphocyte +++ ➤ Eosinophilic polynuclear ++	➤ Diffused ➤ Minimal ➤ Lymphocyte +++ ➤ Eosinophilic polynuclear ++ ➤ Neutrophil polynuclear ++ ➤ Multinucleated giant cells +	➤ Diffused ➤ Minimal ➤ Lymphocyte + ➤ Plasmocyte ++ ➤ Macrophage + ➤ Fibroblast ++
Microthrombi	+++	+++	++
Vascular congestion	+++	+++	+++
Consolidation	45%	55%	60%
Alveolar Fibrosis <i>(Masson Trichome staining)</i>	Negative	Negative	Negative
Co-infection <i>(Periodic Acid Schiff staining)</i>	Negative	Negative	Negative

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DISCUSSION

I. A historic glance at ARDS

In their ground-breaking paper of 1967, Ashbaugh et al. (4) were the first to describe the syndrome that would become one of the distinctive sub-fields of intensive care medicine, namely ARDS. The 12 patients' observations outlined a hypoxemic respiratory failure with patchy bilateral alveolar infiltrates. It also contains a description of the course of the disease, possible causes, and risk factors, pathological findings, loss of surfactant activity, the effectiveness of Positive End-Expiratory Pressure (PEEP) at improving oxygenation, and the conflicting effect of corticosteroids, inotropes, and diuretics. It is noteworthy to mention that, necropsy revealed hyaline membrane and diffuse interstitial inflammation without any sign of micro-thrombi.

In 1994, in the effort to clarify and standardize the concept of Acute Lung Injury (ALI) and ARDS the American-European Consensus Committee was formed (11). ALI was defined as an acute syndrome of inflammation and increased pulmonary permeability that is associated with a constellation of radio-clinical abnormalities, which included ARDS as a severe subgroup. Although both categories demonstrate bilateral infiltrates seen on frontal chest radiograph, the gas exchange impairment in ALI was characterized by $PaO_2/FiO_2 \leq 300$ mm Hg (regardless of PEEP level), whereas ARDS was reduced to $PaO_2/FiO_2 \leq 200$ mm Hg. Therefore, all patients exhibiting ARDS have ALI, but not all patients with ALI have ARDS. The Committee also focused on compartmentalizing risk factors into direct and indirect injuries that induce defects of surfactant balance as well as a ventilation-perfusion mismatch. On one hand, direct insults were portrayed in aspiration, diffuse pulmonary infection; near-drowning, toxic inhalation, and lung contusion, on the other hand, indirect injuries were outlined in sepsis syndrome, hyper-transfusion for emergency resuscitation, and cardiopulmonary bypass.

After 18 years of extensive ARDS research, fundamental concerns have been raised regarding the explicit onset time frame, the sensitivity of PaO_2/FiO_2 to different ventilator settings (PEEP mainly), interpretation variability of the chest radiograph, and difficulties ruling out hydrostatic oedema.

Table III. The AECC Definition —Limitations and Methods to Address These in the Berlin Definition(9)

	AECC definition	AECC limitations	Berlin definition modifications
Timing	Acute onset	No exact timeframe	Acute time onset specified
ALI category	All patients with $P_{aO_2}/F_{iO_2} \leq 300$ mm Hg	Misinterpret as $P_{aO_2}/F_{iO_2} = 201-300$ mm Hg leading to	3 mutually exclusive severity subgroups of ARDS ALI term was removed
Oxygenation	$P_{aO_2}/F_{iO_2} \leq 300$ mm Hg (regardless of PEEP)	Inconsistency of P_{aO_2}/F_{iO_2} ratio due to the effect of PEEP	Minimal PEEP level added across subgroups
Chest radiograph	Bilateral infiltrates observed on frontal image	Wide interpretation variability	Image abnormalities explicitly clarified
Risk Factors	None	Not formally included in the definition	Included When none is identified, need to objectively rule out hydrostatic oedema
Abbreviations: AECC; American European Consensus Conference, ALI; Acute Lung Injury, ARDS; Acute Respiratory Distress Syndrome, F_{iO_2} ; Fraction of inspired oxygen, P_{aO_2} ; arterial partial pressure of oxygen, PEEP; Positive End-Expiratory Pressure.			

Considering that the lack of a standardized terminology hindered clinical research, the Berlin definition was introduced in 2012 to add clarity and uniformity to the notion of ARDS (9). All changes were made with the understanding that syndrome definitions must meet three criteria: practicality, reliability, and validity.

The following update was fundamental because the AECC's definition of ALI as broad-spectrum was withdrawn. The task force suggested 3 mutually exclusive categories of ARDS based on the degree of hypoxemia: mild ($200 \text{ mm Hg} < P_{aO_2}/F_{iO_2} < 300 \text{ mm Hg}$ with PEEP or CPAP $\geq 5 \text{ cm H}_2\text{O}$), moderate ($100 \text{ mm Hg} < P_{aO_2}/F_{iO_2} < 200 \text{ mmHg}$ with PEEP $\geq 5 \text{ cm H}_2\text{O}$), and severe ($P_{aO_2}/F_{iO_2} < 100 \text{ mmHg}$ with PEEP $\geq 5 \text{ cm H}_2\text{O}$).

It is crucial to note that, the onset must occur within one week of recognized clinical abnormalities or the beginning of new or worsening respiratory symptoms. The panel retained bilateral opacities on the chest radiograph consistent with pulmonary oedema as defining criterion, which could not be completely explained by cardiac failure or fluid overload. Moreover,

diffuse alveolar damage was perceived to be the hallmark of the acute phase of ARDS, although not pathognomonic of the disease (12).

Positive end-expiratory pressure can markedly affect PaO₂/FIO₂; therefore, a minimum level of PEEP (5 cm H₂O), which can be delivered noninvasively in mild ARDS, was included in the draft definition of ARDS. A minimum PEEP level of 10 cm H₂O was proposed and empirically evaluated for the severe ARDS category. It is unlikely, for example, that the requirement for a minimal level of PEEP would explain the significant variation in incidence figures.

Although their approach brought uniformity to the definition, it presented a serious challenge in low-income countries. This signifies that ARDS could not be identified especially in settings with scarce resources, considering the requirement of arterial blood gas measurements and chest radiographs. It is reasonable to assume that material shortage challenges diagnosing patients with ARDS, but also prevents physicians from having providing timely adequate medical care.

Moreover, an alternative Kigali definition has been proposed to further simplify the identification of ARDS in settings with scarce access to mechanical ventilation and supplemental oxygen and to adapt its feasibility (13). Furthermore, it was recommended to use SpO₂/FiO₂ rather than PaO₂/ FiO₂ in defining the hypoxemia cut-off and severity of ARDS: SpO₂/FiO₂ ≤ 315 (with a requirement of SpO₂ < 97%). It also went further to replace chest radiograph with lung ultrasound, considering that recent studies validated the utility of lung ultrasound in detecting alveolar filling and consolidation accurately in ARDS(14) (15). As a result, the combination of pulmonary ultrasonography evaluation with SpO₂/FiO₂ measurement provides a relatively sensitive approach for identifying individuals who fulfil standard ARDS oxygenation and imaging criteria (16). It is important to mention that; minimal PEEP requirement was called into question and eventually discarded as a criterion, as a recent paper illustrated the variability of hypoxemia with different PEEP levels and suggests that hypoxemia at lower PEEP levels may be more predictive of death (17).

Table IV. Kigali modification proposal (13).

	Berlin criteria	Kigali modification
Timing	Within 1 week of a known clinical insult or new or worsening respiratory symptoms	Within 1 week of a known clinical insult or new or worsening respiratory symptoms
Oxygenation	$Pa_{O_2}/Fi_{O_2} \leq 300$ mmHg	$Sp_{O_2}/Fi_{O_2} \leq 315$
PEEP requirement	Minimum 5cm H ₂ O PEEP required by invasive mechanical ventilation (non-invasive accepted for mild ARDS)	No PEEP requirement
Chest imaging	Bilateral opacities not fully explained by effusion, lobar /lung collapse, or nodules by chest radiograph or CT scans	Bilateral opacities not fully explained by effusion, lobar /lung collapse, or nodules by lung ultrasound
Origin of oedema	Respiratory failure is not fully explained by cardiac failure nor fluid overload (if no risk is present, need echocardiography to exclude hydrostatic oedema)	Respiratory failure is not fully explained by cardiac failure nor fluid overload (if no risk is present, need echocardiography to exclude hydrostatic oedema)
Abbreviations: PEEP; Positive End-Expiratory Pressure, FiO ₂ ; Fraction of inspired oxygen, PaO ₂ ; arterial partial pressure of oxygen, SpO ₂ ; peripheral capillary oxygen saturation.		

II. C-ARDS

In the early months of 2020, the world witnessed an outbreak of the severe respiratory syndrome coronavirus (SARS-Cov-2), which caused a tremendous flood of coronavirus-related pneumonia. In most cases, coronavirus disease 2019(COVID-19) is rapidly resolved, whereas 26% require intensive care unit admission (18). Clinical manifestations of COVID-19 vary from mild pneumonia (dry cough, fever, fatigue...) to progressive Acute Respiratory Distress Syndrome (ARDS).

Since the beginning of the pandemic, many attempts have been made to understand C-ARDS pathophysiology, and therefore aroused global controversy among clinicians and researchers.

The pivotal question revolved around if acute respiratory failure due to COVID-19 falls into the definition of ARDS, but also reasons behind severe blood-gas exchange deterioration (PaO₂/FiO₂ ratio) compared to non-C-ARDS (19).

Xu Li and Xiaochun Ma (20) claimed that C-ARDS does not fulfil the criteria of a typical ARDS, and argued that the clinical manifestation did not correspond to the severity of the laboratory and radiological images, as well as the time of ARDS onset exceeded 1 week (median was 8-12 days).

Nevertheless, in his cutting edge observation; Gattinoni et al. stated that despite matching the Berlin criteria, C-ARDS exhibits singular features of associating severe hypoxemia with well-maintained respiratory mechanics (21).

Moreover, a fundamental distinction revealed C-ARDS phenotypes; Type 1 (H) with near-normal pulmonary compliance with isolated viral pneumonia and Type 2 (L) with decreased pulmonary compliance (22). In the same report, it has been suggested that Type L, as an early phenotype, is characterized by profound hypoxemia with relatively high compliance (> 50 ml/cmH₂O) paired with high gas volume, thus a minimal percentage of non-aerated tissue and high venous admixture. Whereas in 20-30% of cases, patients present Type H aligned with low respiratory compliance (< 40 mL/ cm H₂O), but high lung weight due to lung diffuse oedema and inflammation. Interestingly, it was affirmed that C-ARDS patients reached, over the course of hospitalisation, a higher value of extravascular lung water index compared to non-C-ARDS which reflects the volume of inflammatory fluid and tissue accumulated during lung injury (19).

On these distinctive grounds, a crucial question rose: How does COVID-19 damage the lungs to cause a rapidly progressive onset of profound hypoxemia? We conducted this Open-Lung-Biopsy in mechanically ventilated patients to identify the pathophysiology of C-ARDS and deduct potential therapeutic targets to eventually allow clinicians to tailor therapy to individuals, making treatment more effective.

III. Pathogenesis

To our knowledge, this is the first report addressing the pulmonary lesions in C-ARDS. In all samples, a diffuse pattern of exudative and early proliferative phases of diffuse alveolar damage was observed. It is noteworthy to mention that, in our report hyperplastic Type 2 pneumocyte with an atypical appearance suggested viral cytopathic changes, thus a direct attack on the lung alveoli. The most striking pattern to emerge from this data was the prominent thrombi in alveolar microvascular beds.

These results correlate favourably with Ackerman et al.(23), and further supports the distinctive angiocentric features of COVID-19. Moreover, the lungs from patients with COVID-19 in the same study had widespread vascular thrombosis with microangiopathy and occlusion of alveolar capillaries. Two hospitals in northern Italy analysed lung tissue samples from 38 patients who succumbed to COVID-19, and demonstrated consistent diffuse alveolar damage, which was observed in all cases, included capillary congestion, interstitial and intra-alveolar oedema, dilated alveolar ducts and collapsed alveoli, hyaline membranes composed of serum proteins and condensed fibrin, and loss of pneumocytes (24). More recent papers highlight the presence of platelet-fibrin thrombi in small arterial vessels as predominant findings in lungs (24-29).

Significant new vessel growth through a mechanism of intussusceptive angiogenesis was noted, and it was hypothesized that a greater degree of endothelialitis and thrombosis in the lungs from patients with COVID-19 may contribute to the relative frequency of angiogenesis (23). Interestingly, it was demonstrated, using Ki67 immunostaining, that vascular endothelium proliferation was frequent and responsible for low perfusion in severe cases of COVID-19 (25).

Other viral respiratory infections have already created substantial public health concerns and led to alarming outbreaks, such as Severe Acute Respiratory Syndrome (SARS), H1N1, and Middle East Respiratory Syndrome-Related Coronavirus (MERS) in 2003, 2009, and 2012 respectively. Table V portrays a pulmonary pathological comparison between different viral pathogens that were responsible for fatal ARDS.

Table V : Distinctive pathologic features in a panel of viruses responsible for ARDS.

Characteristics	SARS	Swine Flu	MERS	COVID-19
Status	First reported in Asia in February. 2003, 8000 people infected, 774 deaths	First reported in Mexico in April 2009 with 201,200 deaths (26)	First reported in Saudi Arabia in September 2012. 2519 people infected, 866 deaths	First reported in Wuhan, China in December 2019. As of 23 October 2021 , 4,955,403 deaths. (27)
Causative virus	SARS-CoV	A/H1N1	MERS-CoV	SARS-CoV-2
Macroscopy	Edematous lungs with increased gross weight and multiple areas of congestion, enlargement of lymph nodes in the pulmonary hila	Edematous lungs with extensive haemorrhagic appearance with an aspect of red hepatisation (28)	Edematous lungs with increased gross weight and multiple areas of congestion	Edematous lungs with increased gross weights, multiple areas of congestion, and pulmonary embolism (29)
Microscopy	Bronchial epithelial denudation, loss of cilia, squamous metaplasia, acute diffuse alveolar damage, and in the late phase acute fibrinous and organizing pneumonia	Acute phase of diffuse alveolar damage, with end-stage organizing pulmonary fibrosis and occasional microthrombi (30)	Exudative diffuse alveolar damage with hyaline membranes, pulmonary oedema, type II pneumocyte hyperplasia, interstitial lymphocytosis, multinucleate syncytial cells cast formation	Diffuse alveolar damage, severe capillary congestion, interstitial mononuclear cell infiltrates, and multinucleated syncytial cells with atypical enlarged pneumocytes, and consistent microthrombosis(31)

IV. Cytokine Storm

For these inpatients, the illness severity spectrum includes not just pneumonia, pulmonary oedema, but also ARDS, which is a cause of death in 70% of fatal COVID-19 cases and is characterised by aggressive inflammatory responses of the host response (32), as exhibited in our case series.

When viral elements of Sars-CoV-2 penetrate lungs, the virus uses angiotensin-converting enzyme II (ACE 2) as host entry through epithelial and endothelial cells, leading to an increased serum level of angiotensin II (Ang II) due to the scarcity of ACE 2 surface expression (33) (34). It is noteworthy to mention that, Ang II appear to function as a vasoconstrictor but also as a pro-inflammatory cytokine via Ang II type 1 receptor (AT1R) (35). Moreover, this AT1R transduces intracellular signalling via complex enzyme and growth factors in order to activate interleukin 6 (IL-6) (36). This chemotaxis cascade of chemokines are known to recruit and migration of leukocytes (macrophages, neutrophils, and T cells) and plasma protein to the inflammation site, where they could eliminate the virus, but simultaneously destruct alveolar epithelial wall, capillary permeability and induce multi-organ failure and ultimately death. This hypothesis is reinforced with our pathological findings of widespread inflammatory infiltrates in the interstitial tissue.

Thus, during extensive lung inflammation in ARDS Ang II-AT1R signalling can generate an IL-6-mediated positive feedback loop, a process known as the IL-6 amplifier resulting in an excessive inflammatory reaction (Fig. 19). The "cytokine storm" is caused by a rapid spike in circulation levels of several pro-inflammatory cytokines such as IL-6, IL-1, TNF-, and interferon (37).

More recent evidence highlights that, when COVID-19 survivors and non-survivors are evaluated, serum level of IL-6 is directly correlated to disease mortality, among the high inflammatory mediators (38) (39). Cytokine storm has been found in a variety of viral diseases, including influenza H5N1 virus, influenza H1N1 virus, and two coronaviruses closely related to COVID-19, SARS-CoV and MERS-CoV (37).

Cytokine storm is a life-threatening complication with a high death rate that necessitates critical care admission and timely-appropriate treatment.

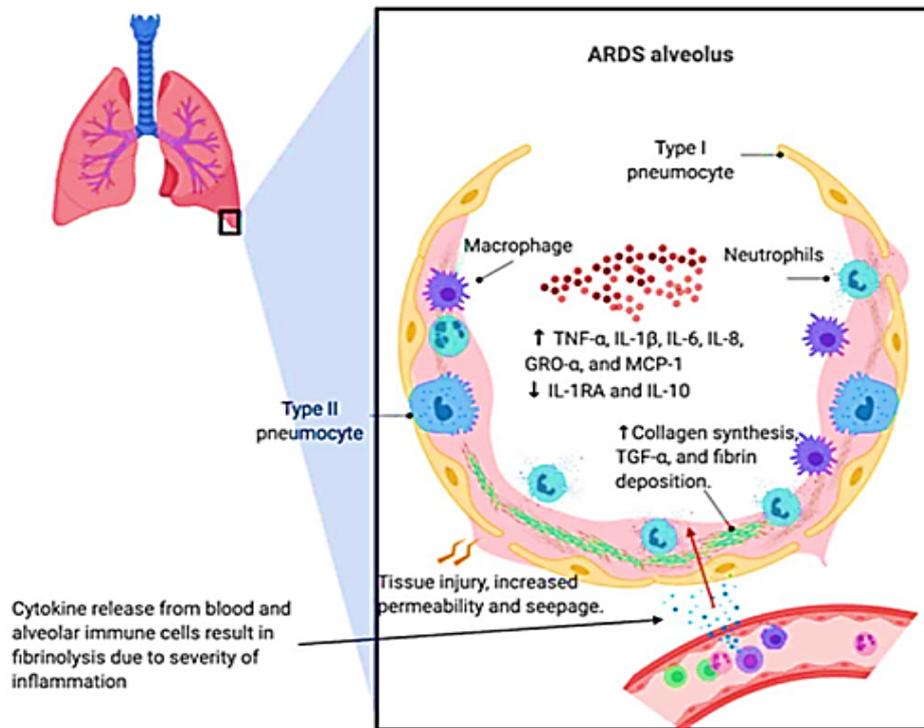


Figure 19: The role of cytokine inducing C-ARDS (40).

V. Immunothrombosis

The presence of SARS-CoV-2 viral elements within the pulmonary and peripheral endothelial cells and the accumulation of inflammatory cells may generate a prothrombotic state by strongly activating endothelial coagulation cascade (41). Moreover, roughly 72% of COVID-19 non-survivors demonstrated evidence of hypercoagulability (42). In addition, accumulating evidence point out that C-ARDS patients may have higher levels of coagulation cascade activation and lower anticoagulant and fibrinolytic system activity than those with ARDS caused by other disorders or illnesses (43) (44). In contrast, ebola, dengue, and other haemorrhagic viruses, which may similarly induce endothelium damage, have been linked to enhanced anticoagulant effects and fatal haemorrhage (45) (46).

Normally, the glycocalyx is fundamental to epithelial and endothelial barrier function under homeostatic conditions and confers an anticoagulant and anti-adhesive surface, however, its shedding is allied with endothelial barrier rupture, and also contribute to increased vascular permeability, leukocyte recruitment and micro-clots in capillaries (47).

It is noteworthy to mention that, endothelial dysfunction is a key component of a variety of disorders, and it also serves as the common denominator for all COVID-19 comorbidities, including hypertension, diabetes, and obesity, all of which are substantial contributors to COVID-19-related mortality (48). On the other hand, endothelial impairment is linked to a low anti-aggregatory prostacyclin production from the endothelial cells and an increased pro-aggregatory thromboxane synthesis from activated platelets (49).

Von Willebrand factor (VWF) levels are significantly elevated in COVID-19 patients (529 U/dL compared to 100 U/dL, normal) further supporting the hypothesis of SARS-CoV-2 induced endothelial dysfunction or damage (50).

VWF is a circulating adhesive glycoprotein released by endothelial cells and platelets, and its levels are raised in disorders such as vasculitis, inflammation, ageing (51), and diabetes (52), all of which are linked to endothelial dysfunction. Furthermore, VWF stimulates platelets, causing them to cluster together (53), functions as a carrier for coagulation factor VIII, and promotes blood coagulation (54).

VWF also has a role in the vasculature system, modulating angiogenesis and vascular permeability.

Interestingly, high inflammation mediated by cytokine, mainly IL-6, (55) regulates coagulation by activating C-reactive protein (CRP), which promotes tissue factor exposure on monocytes and alveolar macrophages (56) and stimulates thrombin production and fibrin deposition.

It is reasonable to assume that the sequestration of endothelial cells-platelet-leukocyte aggregation on the walls of a smaller vessel (Fig. 20) , and the subsequent development of immunothrombosis would be sufficient to cause microthromb in the alveolar-capillary circulation (57) (49), thus a loss of microvascular perfusion in the lungs and other organs

eventually, and in some circumstances the development of disseminated intravascular coagulation (Fig. 21).

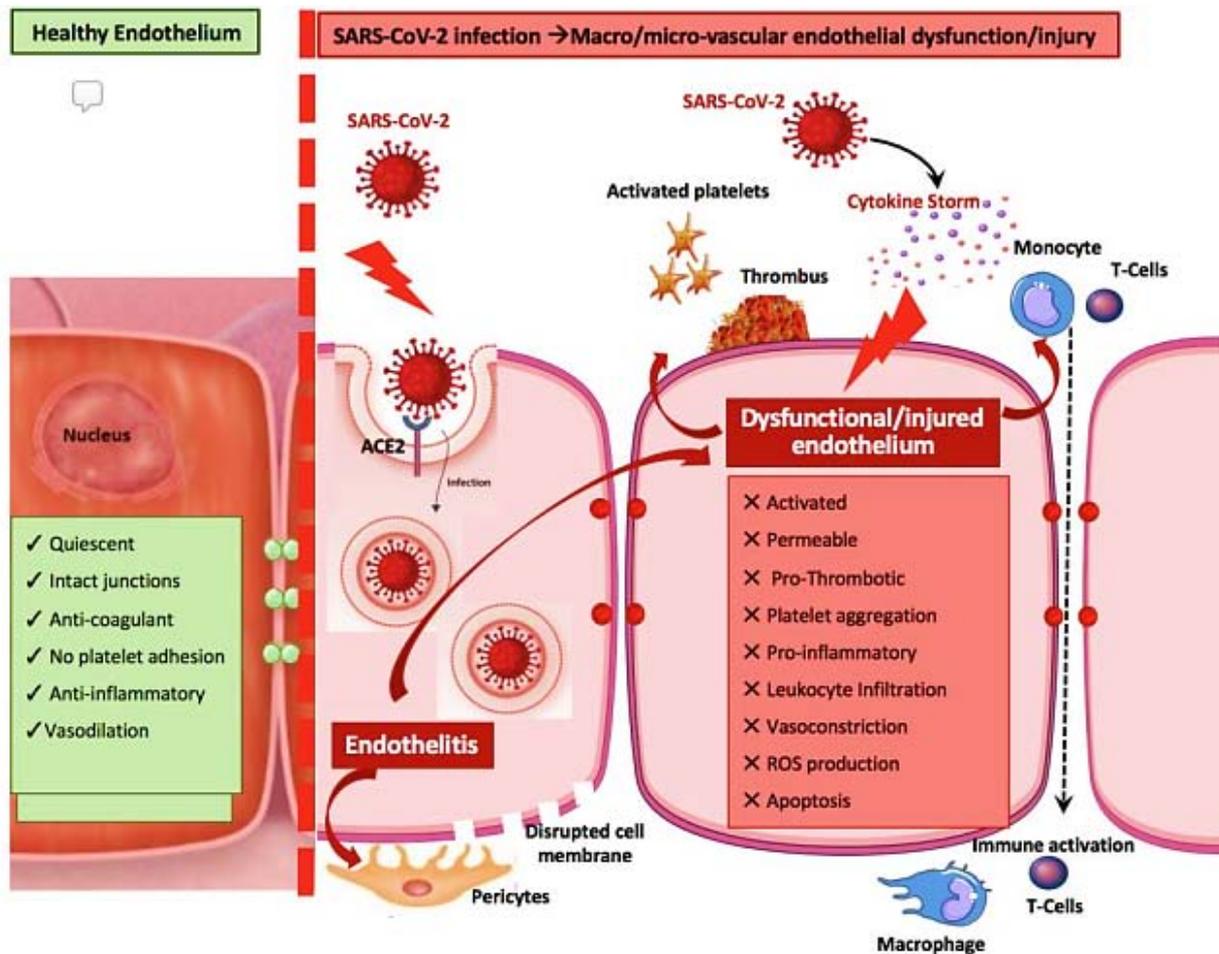


Figure 20: SARS-CoV-2 induced vascular damage (49)

Hypoxia might be, at the same time, the outcome of vascular occlusion and also an accelerator of thrombus formation and progressively exhaust the fibrinolytic activity of the endothelium, acting as a positive loop of reciprocal induction (58).

Additionally, it was noted that mechanical ventilation itself can cause local and systemic inflammatory activation and a hypercoagulable state (59).

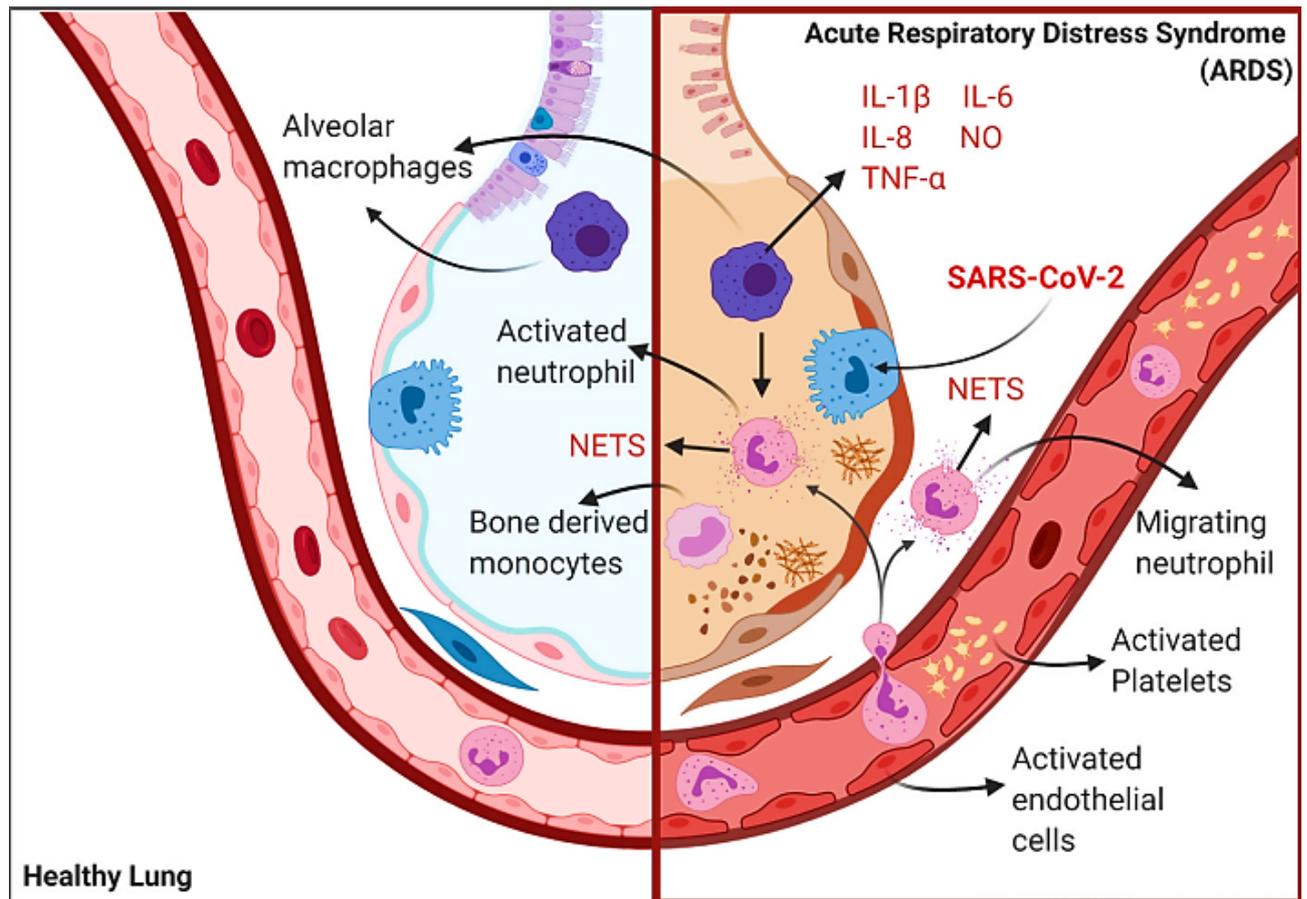


Figure 21: The pathophysiology of COVID-19 ARDS.(60)

Our patients demonstrated elevated levels of D-dimer and it only progressed over time. A meta-analysis of 16 studies found that D-dimer levels were significantly higher in COVID-19 patients compared to healthy controls, in COVID-19 patients with severe disease or a composite end-point compared to non-severe disease, in ARDS patients compared to non-ARDS patients, and in deceased ARDS patients compared to ARDS patients who survived (61).

Ciceri et al. have proposed MicroCLOTS (microvascular COVID-19 lung vessels obstructive thrombo-inflammatory syndrome) as a new name for the severe pulmonary disorder related to COVID-19 (62). They hypothesise that, in predisposed individuals, alveolar viral damage is followed by an inflammatory reaction and by microvascular pulmonary thrombosis.

VI. Co-infection

In our samples, bacterial and fungal cultures were negative, in contrast with the recent finding of bacterial abscesses in 4 patients among 38 who succumbed to COVID-19, with a single fungal abscess in one, and were presumed to be formed after hospital admission (24). SARS-CoV-2 RT PCR was positive in two of our patients, whereas other respiratory pathogens were not present in lung tissue.

Other studies focused on the examination of broncho-alveolar lavages within 24h of tracheal intubation, and interestingly early co-infection was demonstrated in 13 patients out of 47 (63). Three bacterial species accounted for $\geq 90\%$ of all identified bacteria: *Staphylococcus aureus* (all methicillin-sensitive), *Haemophilus influenzae*, and *Streptococcus pneumoniae*.

Extreme caution must be taken interpreting these results, as co-infection with other pathogens cannot be ruled out since it could be present in another pulmonary parenchyma that is not included in the biopsy fragment. It also appears to be difficult to distinguish between morbid infection and tissue colonization.

It would be reasonable to hypothesize, that patients with C-ARDS may be considered as easy prey for secondary infection due to their immune-compromised state, portrayed in lymphopenia in particular.

VII. Therapeutic implications

1. Anticoagulants

Heparin is a polysaccharide that was first extracted from mammalian animal tissue in 1916 (64), its reliance on anti-thrombin to inhibit blood clot formation renders the medicine an indirect antithrombotic agent, and the lack of intrinsic fibrinolytic action inhibits thrombi disintegration after they have formed (65) (66). In addition, heparin is a well-tolerated anticoagulant drug that has been used efficiently for more than 80 years and has few relatively

manageable side effects since it is a natural product (67). Moreover, heparin belongs to a distinct class of medications with effective antidotes, making its use in practice safe.

Besides, heparins also present an interesting immune-modulatory activity: it was found that a non-anticoagulant fraction of enoxaparin was reported as a partial inhibitor of IL-6 and IL-8 release (68). Furthermore, several studies have demonstrated the anti-inflammatory properties of heparin (69) (70), which may be helpful to manage this disease, but also provide endothelium protection.

Our patients showed coagulation abnormalities and received a therapeutic dose of LMWH after their admission to the ICU. Of note, there are several possible explanations for the inefficacy of LMWH administered in our patients. On the one hand, perhaps an asymptomatic pulmonary embolism was already present before the hospitalization of our hypercoagulable patients, considering their advanced age and their cardiovascular comorbidities but also the delay between the symptom onset and their hospitalization.

In line with, a recent major randomized clinical trial that included 1098 patients; proved that in critically ill patients with COVID-19 an initial strategy of therapeutic-dose anticoagulation with heparin did not result in a greater probability of survival to hospital discharge, or a greater number of free organ-support days than did usual-care pharmacologic thrombo-prophylaxis (71).

In another study, severe COVID-19 patients admitted to the ICU were categorised into one of the four groups (fixed-dose, increasing amount, decreasing dose, multiple changes in quantities). As result, no significant difference in dose of anticoagulants between survivors and non-survivors was found and showed no impact in 28-day survival among four strategies of dose modification (72).

In a multicentre randomised clinical trial including COVID-19 unstable patients with elevated D-dimer concentration, a 30-day course of therapeutic anticoagulation (rivaroxaban at 20 mg daily or enoxaparin 1 mg/kg twice daily) did not result in better clinical outcomes, when compared with in-hospital prophylactic anticoagulation with heparin (73). On the contrary, therapeutic anticoagulation led to a higher incidence of bleeding than did in-hospital prophylactic anticoagulation.

It seems plausible that, for anticoagulation to be **efficient** it should be **timely-appropriate**, in parallel with the micro-thrombi formation and elevated D-dimer, before shifting to a clinical unstable status. In contrast with previous studies mentioned above, a large randomized clinical trial in non-critically ill patients with Covid-19 demonstrated that an initial prescription of therapeutic-dose anticoagulation with LMWH improved the probability of survival to hospital discharge with reduced use of organ support, as compared with usual-care thromboprophylaxis (74). Moreover, therapeutic-dose anticoagulation was beneficial regardless of the patient's baseline d-dimer level.

Another anticoagulation approach was suggested, using nebulised unfractionated heparin reaching directly the lung micro-environment. In addition to its anti-viral (75) (76), anti-inflammatory (77) (78) and mucolytic effects (79) nebulised unfractionated heparin targets pulmonary fibrin deposition and inflammation, as well as local administration to the lungs, offers higher doses and enhances local performance, minimises the danger of systemic haemorrhage, and is more beneficial than intravenous treatment (80) (81). It is noteworthy to mention that, pulmonary bleeding did not result from the use of nebulised unfractionated heparin in other respiratory settings (82).

2. Corticosteroids

From another angle, our findings would seem to support the role of corticosteroid therapy. They are steroid hormones that are produced as a result of cholesterol metabolism. It is well known that glucocorticoids reduce endothelial leakage by decreasing capillary permeability and lowering leukocyte migration to the inflammation site, while effectively stopping the inflammatory cascade. It is fundamental to state that; dexamethasone is associated with decreased capillaries permeability, in addition to reduced neutrophil and lymphocyte migration into the inflammatory sites (Fig. 22) (64).

Concerning C-ARDS, it was reported in a single-blind, randomised controlled clinical trial involving 68 severe COVID-19 patients that injecting 250 mg/day of methylprednisolone for 3 days, during the early pulmonary phase lead to a substantial death risk decrease as well as amelioration of ventilation and inflammatory markers (83). Moreover, it was outlined that a high-dose (1000 or 500 mg/day), short-term (3 days) methylprednisolone intravenously enabled extubation of the patients within seven days (84).

The World Health Organization (WHO) have conflicting views concerning the use of corticosteroids in COVID-19 and does not recommend their use in routine practice unless under clinical trial conditions (85). However, it is pragmatic to consider a powerful anti-inflammatory medication, such as dexamethasone therapy, that is required to address the excessive inflammation. Dexamethasone may disrupt the development of the host's natural immunity and abrogate antiviral response, such as in the early stages of the disease, resulting in a delayed viral clearance (64).

The RECOVERY trial investigated the efficacy of dexamethasone in 2104 hospitalized patients (Vs. 4321 standard-care) that received 6 mg once daily for up to 10 days and concluded to reduced 28-day mortality than usual care in patients who were receiving invasive mechanical ventilation or non-invasive ventilation (86). Furthermore, 20 mg of dexamethasone intravenously daily for 5 days, followed by 10 mg of dexamethasone daily for 5 days or until ICU discharge led to a significant increase of survival and free of mechanical ventilation days, in a randomized trial of 299 adults with moderate or severe C-ARDS (87).

It seems reasonable to say that, glucocorticoids use has a clear benefit in the early onset of hypoxemia correlated with inflammatory diffuse alveolar damage, although it might slow viral RNA clearance and increase in antibiotic use and infections without a prolonged hospital stay or increased mortality (88).

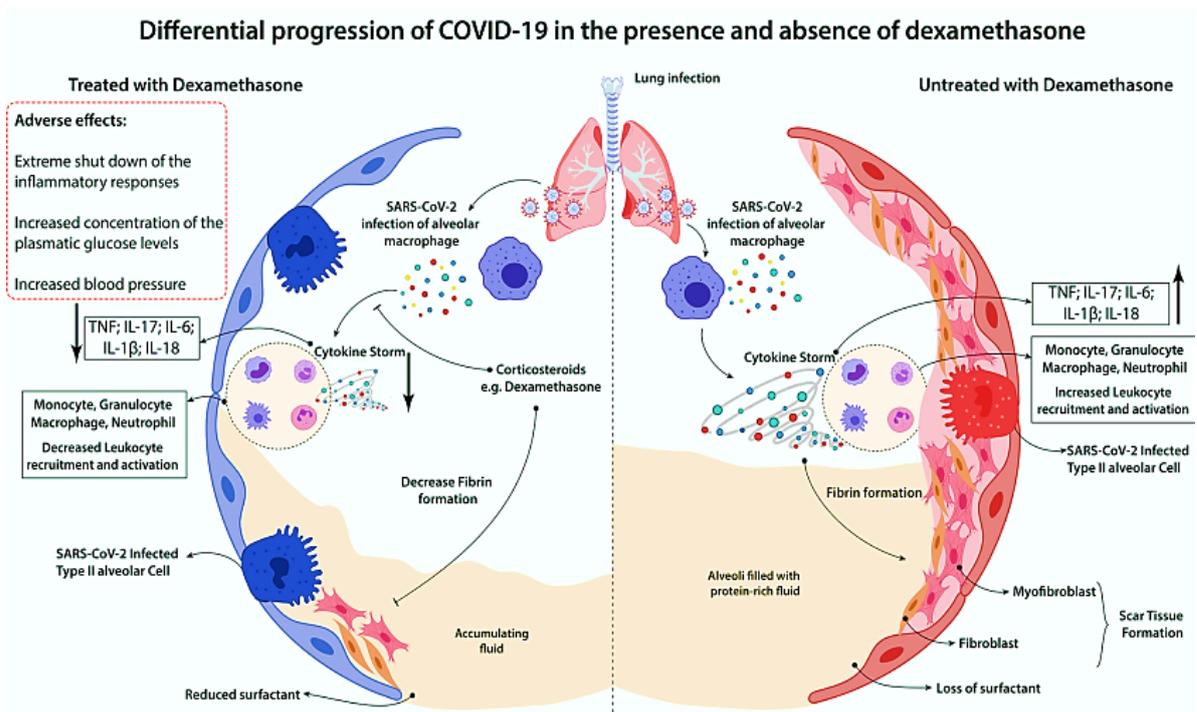


Figure 22: Differential progression of COVID-19 in the presence and absence of dexamethasone. (64)

3. Ventilation management

A key aspect of care should be addressed in ventilation management, not all cases of C-ARDS are considered similar. COVID-19 provides us with a crucial lesson: the broad spectrum of C-ARDS may necessitate alternative ventilatory settings, depending on how they influence lung characteristics: What is a protective tactic in one circumstance may turn out to be a potentially lethal strategy in another (89). Gattinoni et al. conceptualised the L and H type but also differentiated between the distinctive ventilation treatment in his cutting-edge papers (90) (91), as follows (Fig. 23) :

- Initially, we should aim to improve hypoxemia through an increase in FiO₂.
- Several non-invasive alternatives are designed for Type L patients with persistent dyspnea (high-flow nasal cannula, continuous positive airway pressure, or non-invasive ventilation), assuming that they do not make excessive inspiratory efforts.

- If the respiratory drive is not lowered by oxygen delivery and non-invasive support, however, consistently high spontaneous inspiratory attempts increase tissue stresses while also increasing pulmonary transvascular pressures, vascular flows, and fluid leaks.
- Risk of lung injury increases during the transition from the Type L to the Type H phenotype, which could be determined with the magnitude of inspiratory pleural pressures swings.
- Therefore, early intubation, effective sedation, and/or paralysis should be done as soon as possible to break the process.
- Once intubated and deeply sedated, the Type L patients, if hypercapnic, can be ventilated with volumes greater than 6 ml/kg (up to 8-9 ml/kg), as the high compliance results in tolerable strain without the risk of ventilation-induced-lung-injury (VILI).
- Targeting lower PEEP (8-10 cm H₂O) is appropriate since lungs have low recruitability; higher levels will decrease pulmonary compliance and can impact right heart function.
- Prone position may be used as a rescue manoeuvre not to aerate collapsed alveoli, but only to facilitate the redistribution of pulmonary blood flow.
- The type H phenotype progressively develops with the increase of lung oedema and infiltrates. Of note, complete respiratory mechanics assessment performed 4-6 days later in 15 patients suggested that initial status may change more frequently from poorly recruitable to highly recruitable than the reverse (92).
- In this advanced state, it is advisable to apply a more conventional lung-protective strategy: higher PEEP (15 cm H₂O), lower tidal volume (6 mL/kg), and prone positioning while minimizing oxygen consumption.

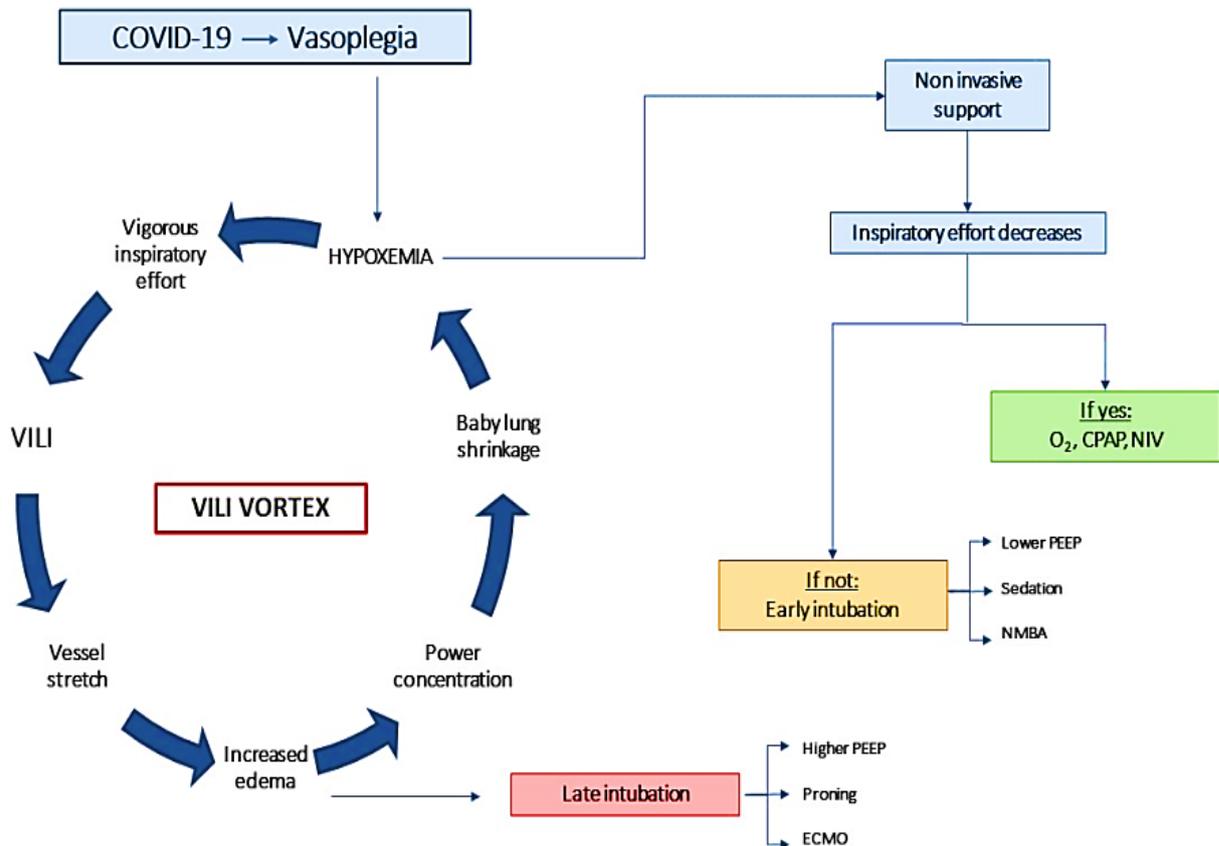


Figure 23: Drivers and Interrupters of Progressive Lung Injury in COVID-19 Infection (93).

VIII. Limitations

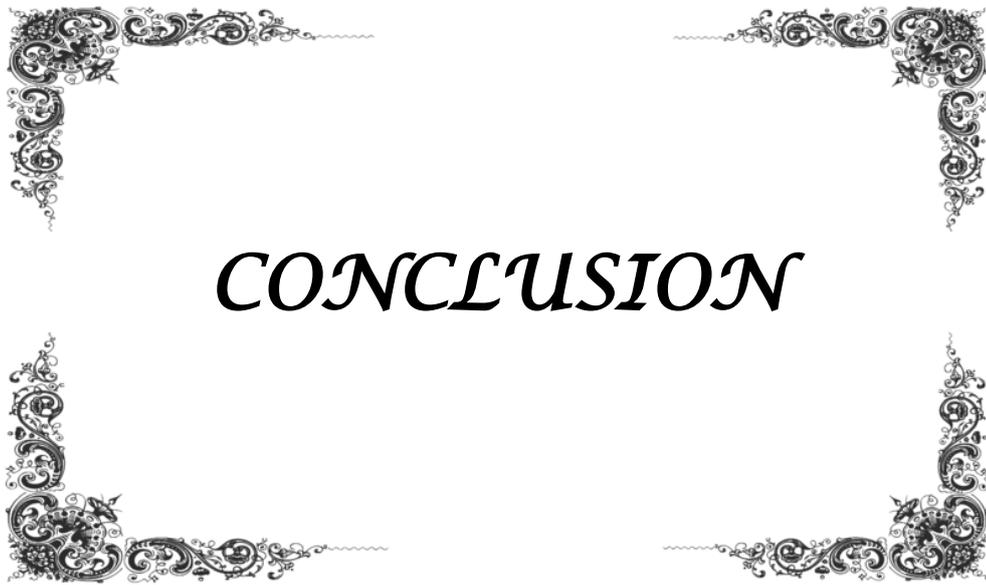
The present study has several limitations; we did not investigate the samples using immune-histochemical staining for additional insights, due to lack of materials. No autopsy was performed to determine the presence of pulmonary embolism in the main vessels.

Despite the sample size, we believe our preliminary work could be a starting point to further provide concrete and substantial answers regarding the mechanism and pathogenesis of severe hypoxemia in C-ARDS and its management.

IX. Strength points

It's noteworthy to mention that our novel work contains some strength points:

- It was a prospective study carried out during the first wave of COVID-19 pandemic that tremendously improved our comprehension of the pathophysiology and also influenced our clinical management.
- To the best of our knowledge, this is the first study in the world and in Morocco to address the pathology in C-ARDS patients.
- It was specifically during the first 24h of intubation to avoid Ventilation induced Lung Injury bias
- Our work was published (94) in Biomed Research International Journal with an impact factor value of 2.5 and 3 other publications have already cited our study including the Lancet Respiratory Journal.



CONCLUSION

The evidence from this study allows us to not only hypothesize that SARS-CoV-2 directly attacks lung alveoli leading to diffuse alveolar damage and more particularly plurifocal fibrin microthrombi in the peripheral vasculature beds, but also elucidate the mechanism behind VA/Q mismatch in severe C-ARDS hypoxemia.

Immunothrombosis may be implicated in the dead space effect through compromising pulmonary perfusion in the early stages, followed by cytokine storm that tends to cause diffuse alveolar damage which is solely responsible for the shunt effect.

Additionally, this research points out the crucial benefit of an early therapeutic dose of LMWH anticoagulant therapy (enoxaparin 1mg/kg twice daily) and corticosteroids in particular dexamethasone (6 mg once daily), in improving survival and lowering C-ARDS mortality.

Despite sample size, we believe that our preliminary report provides a shred of evidence and insight concerning the pathophysiology of C-ARDS. It goes without saying, that further investigation should be conducted in order to determine pertinently C-ARDS hypoxemia mechanisms and the proper ventilation and therapeutic management.



Fiche d'exploitation

IP :

I. Identité :

- Nom et prénom : Sexe : F M
- Âge : Origine : Urbain Semi Urbain Rural Non connu

II. Antécédents :

1. Personnels : Oui Non

1.1. Médicaux :

- HTA Coronaropathie Asthme BPCO Diabète
- Insuffisance rénale C VIH Hémopathie maligne
- Néoplasie Cirrhose Tabac Alcool
- Maladie auto-immune Immunosuppresseurs/CTC
- Autre, Précisez.....
- Traitement actuel :

1.2. Chirurgicaux : Oui Non Précisez :

2. Familiaux : Oui Non Précisez :

III. Symptomatologie

Début de symptômes :

- Fièvre Toux Frissons Dyspnée, Si oui, Stade :
- Fatigue Céphalées Diarrhée Vomissements
- Myalgies Hémoptysie Crachat Douleurs abdominales

IV. Examen physique:

• Fonction respiratoire :

- FR= cpm Cyanose, au niveau de.....
- SLR SpO2= % Auscultation PP =

• Fonction circulatoire

- FC= bpm, PA= mmHg TRC 3s
- Marbrure, au niveau de
- Pâleur Sueurs Auscultation cardiaque=

• **Antibiothérapie– Antiparasitaire–Antiviral**

- Chroloquine 500mgx2/jr OU Plaquenil 200x3/jr
- Azithromycine 250mg/jr
- Ceftriaxone 2g/jr
- Moxifloxacine 400mg/jr
- Lopinavir–Ritonavir
- Favipiravir

• **Anticoagulation**

- Enoxaparine 100 UI/kg x2/jr en SC OU

• **Autre**

- Oméprazole 40mg/jr
- Paracetamol 500mg/6h
- Methylprednisolone 1 mg/kg
- Vitamine C 1g x 2/ jr par SNG
- Sulfate de Zinc 1cp 220mg/jr

• **Ventilation**

- VNI : Oui Non Vent. Invasive : Oui, (tableau) Non

• **Drogues et autres**

- Noradré : Oui Non Dobutamine Oui Non Hémodialyse : Oui Non

VII. Resultat de la biopsie :

1) Analyse anatomopathologique :

2) Analyse microbiologique :

VIII. Evolution

- Sepsis Choc septique Gravité : CURB-65 :..... SOFA :.....
- Insuffisance cardiaque Insuffisance rénale Pneumopathie nosocomiale
- Embolie pulmonaire CIVD Troubles de rythme
- Durée de VNI :J**
- Durée de ventilation mécanique :J**
- Durée d'hospitalisation :.....J** Survivant Décédé, cause :



ABSTRACT

Abstract

Difficulties have risen while managing Acute Respiratory Distress Syndrome (ARDS) caused by COVID-19, although it meets the Berlin definition. Severe hypoxemia with near-normal compliance was noted along with coagulopathy. Understanding the precise pathophysiology of this atypical ARDS will assist researchers and physicians in improving their therapeutic approach.

Previous work is limited to post-mortem studies, while our report addresses patients under protective lung mechanical ventilation. An open-lung minithoracotomy was performed in 3 patients who developed ARDS related to COVID-19 and were admitted to the intensive care unit to carry out a pathological and microbiological analysis on lung tissue biopsy.

Diffused alveolar damage with hyaline membranes was found, as well as plurifocal fibrin microthrombi and vascular congestion in all patients' specimens. Microbiological cultures were negative, whereas qualitative Reversed Transcriptase Polymerase Chain Reaction (RT-PCR) detected SARS-CoV-2 in the pulmonary parenchyma and pleural fluid in two patients.

COVID-19 causes progressive ARDS with onset of severe hypoxemia, underlying a dual mechanism: shunt effect through diffused alveolar damage and dead space effect through thrombotic injuries in microvascular beds. It seems reasonable to manage this ventilation-perfusion ratio mismatch using a high dose of anticoagulant combined with glucocorticoids.

It's noteworthy to mention that our work was published on 10th December 2020, in "Biomed Research International" Journal with an impact factor value of 2.5 and 3 other publications have already cited our study including the **Lancet Respiratory** Journal.

Resumé

Des difficultés sont apparues lors de la gestion du syndrome de détresse respiratoire aiguë (SDRA) causé par la COVID-19, bien qu'il réponde à la définition de Berlin. Une hypoxémie sévère avec une compliance quasi normale a été constatée ainsi qu'une coagulopathie. La compréhension de la physiopathologie précise de ce SDRA atypique aidera les chercheurs et les médecins à améliorer leur approche thérapeutique.

Les travaux antérieurs se limitent à des études post-mortem, tandis que notre rapport porte sur des patients sous ventilation mécanique . Une biopsie à mini-thoracotomie a été réalisée chez 3 patients qui ont développé un SDRA lié au COVID-19 et ont été admis dans l'unité de soins intensifs pour effectuer une analyse pathologique et microbiologique du tissu pulmonaire.

Des dommages alvéolaires diffusés avec des membranes hyalines ont été trouvées, ainsi que des microthrombi de fibrines plurifocales et une congestion vasculaire dans les échantillons de tous les patients. Les cultures microbiologiques se sont révélées négatives, tandis que l'amplification en chaîne par polymérase à transcriptase inverse (RT-PCR) qualitative a détecté le SARS-CoV-2 dans le parenchyme pulmonaire et le liquide pleural de deux patients.

La COVID-19 provoque un SDRA progressif avec installation d'une hypoxémie sévère, à travers un double mécanisme : effet de shunt par des lésions alvéolaires diffusées et effet espace mort par des lésions thrombotiques dans les lits microvasculaires. Il semble raisonnable de gérer cette inadéquation du rapport ventilation-perfusion en utilisant une forte dose d'anticoagulant associée à des glucocorticoïdes.

Il est important à noter que notre travail a été publié le 10 décembre 2020 dans le Journal « **Biomed Research International** » avec une valeur de facteur d'impact de 2,5 ainsi que, 3 autres publications ont déjà cité notre étude, dont le **Lancet Respiratory Journal**.

ملخص

ازدادت الصعوبات خلال ادارة جائحة كوفيد-19 المرتبطة بمتلازمة الضائقة التنفسية الحادة الناجمة عن التعفن الفيروسي سارس-كوف-2 , على الرغم من سهولة تشخيص المرض وفق تعريف برلين لعام 2012 , لوحظ تناقض كبير من حيث الخطورة بين الحالة السريرية للمرضى و النقص الحاد لنسبة تشبع الأنسجة بالأكسجين.

أمام تفاقم الوضعية الوبائية و عدم نجاعة التوصيات العلمية- نسبيا آنذاك- لعلاج هاته المتلازمة غير النمطية و غير المألوفة في أقسام الانعاش.

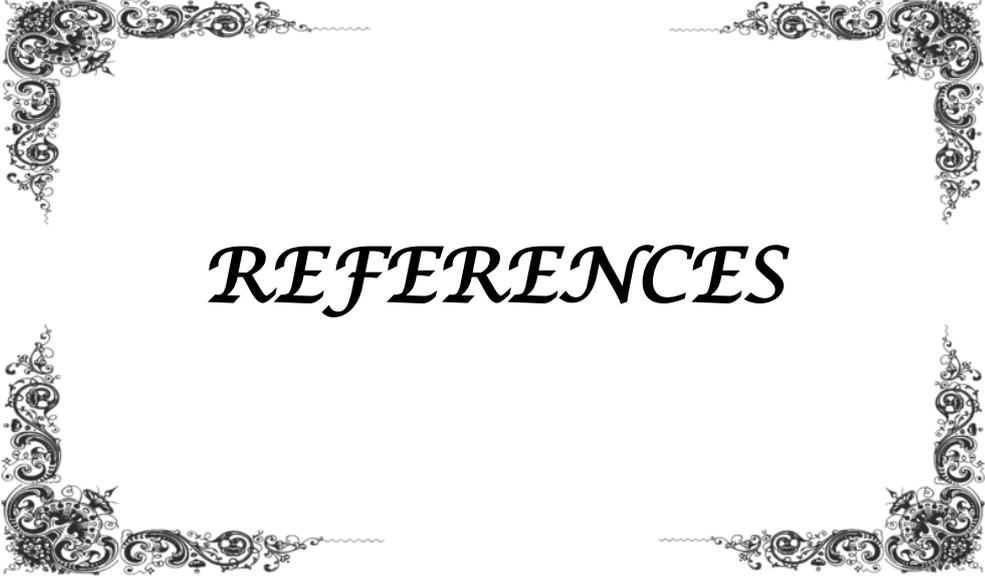
ارتأى الفريق الطبي فهم كيفية تأثير فيروس كورونا-19 المستجد على النسيج الرئوي بغية تحسين النهج العلاجي. اقتصرت الدراسات السابقة على التشريح المرضي لعينات الأنسجة الرئوية للمرضى المتوفين, بينما استهدف هذا البحث العلمي اخذ عينات ثلاث مرضى خاضعين للتنفس الاصطناعي بعد تدهور حالتهم الصحية الحرجة , و ذلك عن طريق عملية جراحية محدودة مع مراعاة الشروط الوقائية لتفادي العدوى.

كشفت النتائج على انتشار تلف الحويصلات الرئوية مع توضع أغشية الهياطين , و كذلك احتقان و تخثر الأوعية الدموية في جميع عينات المرضى. الى جانب ذلك, لم يتم العثور على تعفن بكتيري , و لا فيروسي و لا طفيلي , في حين تفاعل البوليميراز التسلسلي المعكوس للساسرس كوف-2 كان ايجابيا عند مريضين.

يتسبب مرض كوفيد-19 في الإصابة بمتلازمة الضائقة التنفسية الحادة مع ظهور نقص تدريجي في نسبة الأكسجين الدموي الحاد , و يرجع ذلك لآليتين **shunt effect**: من خلال انتشار الاضرار السنخية الى جانب **dead space effect** بواسطة تجلط الدم المجهري في شعيرات الدموية الرئوية. يبدو منطقيا في هذا الاطار استخدام جرعة عالية من مضادات التخثر بالاضافة الى گلوكوكورتيكويد.

من الجدير بالذكر أن عملنا نُشر في 10 ديسمبر 2020 ، في مجلة " Biomed Research

"International مع قيمة عامل التأثير 2.5 وقد استشهدت 3 منشورات أخرى بالفعل بدراستنا بما في ذلك مجلة Lancet Respiratory Journal.

A decorative border consisting of four ornate, symmetrical floral corner pieces arranged in a square pattern around the central text.

REFERENCES

1. **Parsa-Parsi RW.**
The Revised Declaration of Geneva: A Modern-Day Physician's Pledge. *JAMA*. 2017 Nov 28;318(20):1971-2.
2. **Wang D, Hu B, Hu C, Zhu F, Liu X, Zhang J, et al.**
Clinical Characteristics of 138 Hospitalized Patients With 2019 Novel Coronavirus-Infected Pneumonia in Wuhan, China. *JAMA*. 2020 Mar 17;323(11):1061-9.
3. **Gattinoni L, Coppola S, Cressoni M, Busana M, Rossi S, Chiumello D.**
COVID-19 Does Not Lead to a 'Typical' Acute Respiratory Distress Syndrome. *Am J Respir Crit Care Med*. 2020 May 15;201(10):1299-300.
4. **Ashbaugh DG, Bigelow DB, Levine BE.**
ACUTE RESPIRATORY DISTRESS IN ADULTS. :5.
5. **Thompson BT, Guérin C, Esteban A.**
Should ARDS be renamed diffuse alveolar damage? *Intensive Care Med*. 2016 May;42(5):653-5.
6. **Puelles VG, Lütgehetmann M, Lindenmeyer MT, Sperhake JP, Wong MN, Allweiss L, et al.**
Multiorgan and Renal Tropism of SARS-CoV-2. *N Engl J Med*. 2020 Aug 6;383(6):590-2.
7. **Zhang Y, Xiao M, Zhang S, Xia P, Cao W, Jiang W, et al.**
Coagulopathy and Antiphospholipid Antibodies in Patients with Covid-19. *N Engl J Med*. 2020 Apr 23;382(17):e38.
8. **Escher R, Breakey N, Lämmle B.**
Severe COVID-19 infection associated with endothelial activation. *Thromb Res*. 2020 Jun;190:62.
9. **ARDS Definition Task Force, Ranieri VM, Rubenfeld GD, Thompson BT, Ferguson ND, Caldwell E, et al.**
Acute respiratory distress syndrome: the Berlin Definition. *JAMA*. 2012 Jun 20;307(23):2526-33.
10. **Kim D, Quinn J, Pinsky B, Shah NH, Brown I.**
Rates of Co-infection Between SARS-CoV-2 and Other Respiratory Pathogens. *JAMA*. 2020 May 26;323(20):2085-6.
11. **Bernard GR, Artigas A, Brigham KL, Carlet J, Falke K, Hudson L, et al.**
The American-European Consensus Conference on ARDS. Definitions, mechanisms, relevant outcomes, and clinical trial coordination. *Am J Respir Crit Care Med*. 1994 Mar;149(3):818-24.

12. **Katzenstein A-LA, Bloor CM, Leibow AA.**
Diffuse Alveolar Damage-The Role of Oxygen, Shock, and Related Factors. 1976;85(1):20.
13. **Riviello ED, Buregeya E, Twagirumugabe T.**
Diagnosing acute respiratory distress syndrome in resource limited settings: the Kigali modification of the Berlin definition. *Curr Opin Crit Care*. 2017 Feb;23(1):18-23.
14. **Lichtenstein DA, Mezière GA.**
Relevance of lung ultrasound in the diagnosis of acute respiratory failure: the BLUE protocol. *Chest*. 2008 Jul;134(1):117-25.
15. **Lichtenstein D, Goldstein I, Mourgeon E, Cluzel P, Grenier P, Rouby J-J.**
Comparative diagnostic performances of auscultation, chest radiography, and lung ultrasonography in acute respiratory distress syndrome. *Anesthesiology*. 2004 Jan;100(1):9-15.
16. **Bass CM, Sajed DR, Adedipe AA, West TE.**
Pulmonary ultrasound and pulse oximetry versus chest radiography and arterial blood gas analysis for the diagnosis of acute respiratory distress syndrome: a pilot study. *Crit Care Lond Engl*. 2015 Jul 21;19:282.
17. **Caironi P, Carlesso E, Cressoni M, Chiumello D, Moerer O, Chiurazzi C, et al.**
Lung recruitability is better estimated according to the Berlin definition of acute respiratory distress syndrome at standard 5 cm H₂O rather than higher positive end-expiratory pressure: a retrospective cohort study. *Crit Care Med*. 2015 Apr;43(4):781-90.
18. **Wang D, Hu B, Hu C, Zhu F, Liu X, Zhang J, et al.**
Clinical Characteristics of 138 Hospitalized Patients With 2019 Novel Coronavirus-Infected Pneumonia in Wuhan, China. *JAMA*. 2020 Mar 17;323(11):1061-9.
19. **Shi R, Lai C, Teboul J-L, Dres M, Moretto F, De Vita N, et al.**
COVID-19 ARDS is characterized by higher extravascular lung water than non-COVID-19 ARDS: the PiCCOVID study. *Crit Care*. 2021 Dec;25(1):186.
20. **Li X, Ma X.**
Acute respiratory failure in COVID-19: is it "typical" ARDS? *Crit Care*. 2020 Dec;24(1):198.
21. **Gattinoni L, Coppola S, Cressoni M, Busana M, Rossi S, Chiumello D.**
COVID-19 Does Not Lead to a "Typical" Acute Respiratory Distress Syndrome. *Am J Respir Crit Care Med*. 2020 May 15;201(10):1299-300.

22. **Gattinoni L, Chiumello D, Rossi S.**
COVID-19 pneumonia: ARDS or not? *Crit Care*. 2020 Dec;24(1):154, s13054-020-02880-z.
23. **Ackermann M, Verleden SE, Kuehnel M, Haverich A, Welte T, Laenger F, et al.**
Pulmonary Vascular Endothelialitis, Thrombosis, and Angiogenesis in Covid-19. *N Engl J Med*. 2020;9.
24. **Carsana L, Sonzogni A, Nasr A, Rossi RS, Pellegrinelli A, Zerbi P, et al.**
Pulmonary post-mortem findings in a series of COVID-19 cases from northern Italy: a two-centre descriptive study. *Lancet Infect Dis*. 2020 Oct;20(10):1135-40.
25. **Pérez-Mies B, Gómez-Rojo M, Carretero-Barrio I, Bardi T, Benito A, García-Cosío M, et al.**
Pulmonary vascular proliferation in patients with severe COVID-19: an autopsy study. *Thorax*. 2021 Oct;76(10):1044-6.
26. **Dawood FS, Iuliano AD, Reed C, Meltzer MI, Shay DK, Cheng P-Y, et al.**
Estimated global mortality associated with the first 12 months of 2009 pandemic influenza A H1N1 virus circulation: a modelling study. *Lancet Infect Dis*. 2012 Sep;12(9):687-95.
27. **COVID Live Update: 243,851,805 Cases and 4,955,454 Deaths from the Coronavirus – Worldometer [Internet]. [cited 2021 Oct 23].**
Available from: <https://www.worldometers.info/coronavirus/>
28. **Calore EE, Uip DE, Perez NM.**
Pathology of the swine-origin influenza A (H1N1) flu. *Pathol – Res Pract*. 2011 Feb 15;207(2):86-90.
29. **Wichmann D, Sperhake J-P, Lütgehetmann M, Steurer S, Edler C, Heinemann A, et al.**
Autopsy Findings and Venous Thromboembolism in Patients With COVID-19: A Prospective Cohort Study. *Ann Intern Med*. 2020 Aug 18;173(4):268-77.
30. **Hariri LP, North CM, Shih AR, Israel RA, Maley JH, Villalba JA, et al.**
Lung Histopathology in Coronavirus Disease 2019 as Compared With Severe Acute Respiratory Syndrome and H1N1 Influenza. *Chest*. 2021 Jan;159(1):73-84.
31. **Fox SE, Akmatbekov A, Harbert JL, Li G, Quincy Brown J, Vander Heide RS.**
Pulmonary and cardiac pathology in African American patients with COVID-19: an autopsy series from New Orleans. *Lancet Respir Med*. 2020 Jul;8(7):681-6.
32. **Huang C, Wang Y, Li X, Ren L, Zhao J, Hu Y, et al.**
Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China. *Lancet Lond Engl*. 2020 Feb 15;395(10223):497-506.

33. **Kuba K, Imai Y, Rao S, Gao H, Guo F, Guan B, et al.**
A crucial role of angiotensin converting enzyme 2 (ACE2) in SARS coronavirus-induced lung injury. *Nat Med.* 2005 Aug;11(8):875-9.
34. **Verdecchia P, Cavallini C, Spanevello A, Angeli F.**
The pivotal link between ACE2 deficiency and SARS-CoV-2 infection. *Eur J Intern Med.* 2020 Jun;76:14-20.
35. **Eguchi S, Kawai T, Scalia R, Rizzo V.**
Understanding Angiotensin II Type 1 Receptor Signaling in Vascular Pathophysiology. *Hypertens Dallas Tex* 1979. 2018 May;71(5):804-10.
36. **Murakami M, Kamimura D, Hirano T.**
Pleiotropy and Specificity: Insights from the Interleukin 6 Family of Cytokines. *Immunity.* 2019 Apr 16;50(4):812-31.
37. **Ragab D, Salah Eldin H, Taeimah M, Khattab R, Salem R.**
The COVID-19 Cytokine Storm; What We Know So Far. *Front Immunol.* 2020 Jun 16;11:1446.
38. **Zhou F, Yu T, Du R, Fan G, Liu Y, Liu Z, et al.**
Clinical course and risk factors for mortality of adult inpatients with COVID-19 in Wuhan, China: a retrospective cohort study. *Lancet Lond Engl.* 2020 Mar 28;395(10229):1054-62.
39. **Liu B, Li M, Zhou Z, Guan X, Xiang Y.**
Can we use interleukin-6 (IL-6) blockade for coronavirus disease 2019 (COVID-19)-induced cytokine release syndrome (CRS)? *J Autoimmun.* 2020 Jul;111:102452.
40. **McGonagle D, Sharif K, O'Regan A, Bridgewood C.**
The Role of Cytokines including Interleukin-6 in COVID-19 induced Pneumonia and Macrophage Activation Syndrome-Like Disease. *Autoimmun Rev.* 2020 Jun;19(6):102537.
41. **Varga Z, Flammer AJ, Steiger P, Haberecker M, Andermatt R, Zinkernagel AS, et al.**
Endothelial cell infection and endotheliitis in COVID-19. *Lancet Lond Engl.* 2020 May 2;395(10234):1417-8.
42. **Tang N.**
Response to 'Lupus anticoagulant is frequent in patients with Covid-19' (JTH-2020-00483). *J Thromb Haemost JTH.* 2020 Aug;18(8):2065-6.

43. **Ranucci M, Ballotta A, Di Dedda U, Baryshnikova E, Dei Poli M, Resta M, et al.**
The procoagulant pattern of patients with COVID-19 acute respiratory distress syndrome. *J Thromb Haemost.* 2020;18(7):1747-51.
44. **Tang X, Du R-H, Wang R, Cao T-Z, Guan L-L, Yang C-Q, et al.**
Comparison of Hospitalized Patients With ARDS Caused by COVID-19 and H1N1. *Chest.* 2020 Jul 1;158(1):195-205.
45. **Schnittler HJ, Mahner F, Drenckhahn D, Klenk HD, Feldmann H.**
Replication of Marburg virus in human endothelial cells. A possible mechanism for the development of viral hemorrhagic disease. *J Clin Invest.* 1993 Apr 1;91(4):1301-9.
46. **Mahanty S, Bray M.**
Pathogenesis of filoviral haemorrhagic fevers. *Lancet Infect Dis.* 2004 Aug 1;4(8):487-98.
47. **Robba C, Battaglini D, Ball L, Valbusa A, Porto I, Della Bona R, et al.**
Coagulative Disorders in Critically Ill COVID-19 Patients with Acute Distress Respiratory Syndrome: A Critical Review. *J Clin Med.* 2021 Jan 3;10(1):140.
48. **Amraei R, Rahimi N.**
COVID-19, Renin-Angiotensin System and Endothelial Dysfunction. *Cells.* 2020 Jul 9;9(7):1652.
49. **Evans PC, Rainger GE, Mason JC, Guzik TJ, Osto E, Stamataki Z, et al.**
Endothelial dysfunction in COVID-19: a position paper of the ESC Working Group for Atherosclerosis and Vascular Biology, and the ESC Council of Basic Cardiovascular Science. *Cardiovasc Res.* 2020 Dec 1;116(14):2177-84.
50. **Panigada M, Bottino N, Tagliabue P, Grasselli G, Novembrino C, Chantarangkul V, et al.**
Hypercoagulability of COVID-19 patients in intensive care unit: A report of thromboelastography findings and other parameters of hemostasis. *J Thromb Haemost JTH.* 2020 Jul;18(7):1738-42.
51. **Haverkate F, Thompson SG, Duckert F.**
Haemostasis factors in angina pectoris; relation to gender, age and acute-phase reaction. Results of the ECAT Angina Pectoris Study Group. *Thromb Haemost.* 1995 Apr;73(4):561-7.
52. **Stehouwer CD, Nauta JJ, Zeldenrust GC, Hackeng WH, Donker AJ, den Ottolander GJ.**
Urinary albumin excretion, cardiovascular disease, and endothelial dysfunction in non-insulin-dependent diabetes mellitus. *Lancet Lond Engl.* 1992 Aug 8;340(8815):319-23.

53. **Löf A, Müller JP, Brehm MA.**
A biophysical view on von Willebrand factor activation. *J Cell Physiol.* 2018 Feb;233(2):799–810.
54. **Butera D, Passam F, Ju L, Cook KM, Woon H, Aponte-Santamaría C, et al.**
Autoregulation of von Willebrand factor function by a disulfide bond switch. *Sci Adv.* 2018 Feb;4(2):eaq1477.
55. **Hojyo S, Uchida M, Tanaka K, Hasebe R, Tanaka Y, Murakami M, et al.**
How COVID-19 induces cytokine storm with high mortality. *Inflamm Regen.* 2020 Dec;40(1):37.
56. **Xue M, Sun Z, Shao M, Yin J, Deng Z, Zhang J, et al.**
Diagnostic and prognostic utility of tissue factor for severe sepsis and sepsis-induced acute lung injury. *J Transl Med.* 2015 May 30;13:172.
57. **Pfeiler S, Massberg S, Engelmann B.**
Biological basis and pathological relevance of microvascular thrombosis. *Thromb Res.* 2014 May 1;133:S35–7.
58. **Gupta N, Zhao Y-Y, Evans CE.**
The stimulation of thrombosis by hypoxia. *Thromb Res.* 2019 Sep;181:77–83.
59. **Slutsky AS, Ranieri VM.**
Ventilator-induced lung injury. *N Engl J Med.* 2013 Nov 28;369(22):2126–36.
60. **Morris G, Bortolasci CC, Puri BK, Olive L, Marx W, O’Neil A, et al.**
Preventing the development of severe COVID-19 by modifying immunothrombosis. *Life Sci.* 2021 Jan;264:118617.
61. **Vidali S, Morosetti D, Cossu E, Luisi MLE, Pancani S, Semeraro V, et al.**
D-dimer as an indicator of prognosis in SARS-CoV-2 infection: a systematic review. *ERJ Open Res.* 2020 Apr;6(2):00260–2020.
62. **Ciceri F, Beretta L, Scandroglio AM, Colombo S, Landoni G, Ruggeri A, et al.**
Microvascular COVID-19 lung vessels obstructive thromboinflammatory syndrome (MicroCLOTS): an atypical acute respiratory distress syndrome working hypothesis. *Crit Care Resusc J Australas Acad Crit Care Med.* 2020 Apr 15;22(2):95–7.
63. **Kreitmann L, Monard C, Dauwalder O, Simon M, Argaud L.**
Early bacterial co-infection in ARDS related to COVID-19. *Intensive Care Med.* 2020 Sep;46(9):1787–9.

64. **Braz-de-Melo HA, Faria SS, Pasquarelli-do-Nascimento G, Santos I de O, Kobinger GP, Magalhães KG.**
The Use of the Anticoagulant Heparin and Corticosteroid Dexamethasone as Prominent Treatments for COVID-19. *Front Med.* 2021 Apr 23;8:615333.
65. **Oduah EI, Linhardt RJ, Sharfstein ST.**
Heparin: Past, Present, and Future. *Pharm Basel Switz.* 2016 Jul 4;9(3):E38.
66. **Alquwaizani M, Buckley L, Adams C, Fanikos J.**
Anticoagulants: A Review of the Pharmacology, Dosing, and Complications. *Curr Emerg Hosp Med Rep.* 2013 Jun;1(2):83-97.
67. **Gaertner F, Massberg S.**
Blood coagulation in immunothrombosis-At the frontline of intravascular immunity. *Semin Immunol.* 2016 Dec;28(6):561-9.
68. **Shastri MD, Stewart N, Horne J, Peterson GM, Gueven N, Sohal SS, et al.**
In-vitro suppression of IL-6 and IL-8 release from human pulmonary epithelial cells by non-anticoagulant fraction of enoxaparin. *PLoS One.* 2015;10(5):e0126763.
69. **Esmon CT.**
Targeting factor Xa and thrombin: impact on coagulation and beyond. *Thromb Haemost.* 2014 Apr 1;111(4):625-33.
70. **Mousavi S, Moradi M, Khorshidahmad T, Motamedi M.**
Anti-Inflammatory Effects of Heparin and Its Derivatives: A Systematic Review. *Adv Pharmacol Sci.* 2015;2015:507151.
71. **The REMAP-CAP, ACTIV-4a, and ATTACC Investigators.**
Therapeutic Anticoagulation with Heparin in Critically Ill Patients with Covid-19. *N Engl J Med.* 2021 Aug 26;385(9):777-89.
72. **Nadeem R, Thomas SJ, Fathima Z, Palathinkal AS, Alkilani YE, Dejan EA, et al.**
Pattern of anticoagulation prescription for patients with Covid-19 acute respiratory distress syndrome admitted to ICU. Does it impact outcome? *Heart Lung.* 2021 Jan;50(1):1-5.
73. **Lopes RD, de Barros e Silva PGM, Furtado RHM, Macedo AVS, Bronhara B, Damiani LP, et al.**
Therapeutic versus prophylactic anticoagulation for patients admitted to hospital with COVID-19 and elevated D-dimer concentration (ACTION): an open-label, multicentre, randomised, controlled trial. *The Lancet.* 2021 Jun;397(10291):2253-63.

74. **The ATTACC, ACTIV-4a, and REMAP-CAP Investigators.**
Therapeutic Anticoagulation with Heparin in Noncritically Ill Patients with Covid-19. *N Engl J Med.* 2021 Aug 26;385(9):790-802.
75. **Idänpään-Heikkilä I, Simon PM, Zopf D, Vullo T, Cahill P, Sokol K, et al.**
Oligosaccharides interfere with the establishment and progression of experimental pneumococcal pneumonia. *J Infect Dis.* 1997 Sep;176(3):704-12.
76. **Bryan R, Feldman M, Jawetz SC, Rajan S, DiMango E, Tang HB, et al.**
The effects of aerosolized dextran in a mouse model of *Pseudomonas aeruginosa* pulmonary infection. *J Infect Dis.* 1999 Jun;179(6):1449-58.
77. **Camprubí-Rimblas M, Guillamat-Prats R, Lebouvier T, Bringué J, Chimenti L, Iglesias M, et al.**
Role of heparin in pulmonary cell populations in an in-vitro model of acute lung injury. *Respir Res.* 2017 May 10;18(1):89.
78. **Chimenti L, Camprubí-Rimblas M, Guillamat-Prats R, Gomez MN, Tijero J, Blanch L, et al.**
Nebulized Heparin Attenuates Pulmonary Coagulopathy and Inflammation through Alveolar Macrophages in a Rat Model of Acute Lung Injury. *Thromb Haemost.* 2017 Nov;117(11):2125-34.
79. **Tang XX, Ostedgaard LS, Hoegger MJ, Moninger TO, Karp PH, McMenimen JD, et al.**
Acidic pH increases airway surface liquid viscosity in cystic fibrosis. *J Clin Invest.* 2016 Mar 1;126(3):879-91.
80. **Tuinman PR, Dixon B, Levi M, Juffermans NP, Schultz MJ.**
Nebulized anticoagulants for acute lung injury – a systematic review of preclinical and clinical investigations. *Crit Care Lond Engl.* 2012 Dec 12;16(2):R70.
81. **Camprubí-Rimblas M, Tantinyà N, Bringué J, Guillamat-Prats R, Artigas A.**
Anticoagulant therapy in acute respiratory distress syndrome. *Ann Transl Med.* 2018 Jan;6(2):36.
82. **Mulloy B, Hogwood J, Gray E, Lever R, Page CP.**
Pharmacology of Heparin and Related Drugs. *Pharmacol Rev.* 2016 Jan;68(1):76-141.
83. **Edalatifard M, Akhtari M, Salehi M, Naderi Z, Jamshidi A, Mostafaei S, et al.**
Intravenous methylprednisolone pulse as a treatment for hospitalised severe COVID-19 patients: results from a randomised controlled clinical trial. *Eur Respir J.* 2020 Dec;56(6):2002808.

84. **So C, Ro S, Murakami M, Imai R, Jinta T.**
High-dose, short-term corticosteroids for ARDS caused by COVID-19: a case series. *Respirol Case Rep.* 2020 Aug;8(6):e00596.
85. **Jin Y-H, Cai L, Cheng Z-S, Cheng H, Deng T, Fan Y-P, et al.**
A rapid advice guideline for the diagnosis and treatment of 2019 novel coronavirus (2019-nCoV) infected pneumonia (standard version). *Mil Med Res.* 2020 Feb 6;7(1):4.
86. **The RECOVERY Collaborative Group.**
Dexamethasone in Hospitalized Patients with Covid-19. *N Engl J Med.* 2021 Feb 25;384(8):693-704.
87. **Tomazini BM, Maia IS, Cavalcanti AB, Berwanger O, Rosa RG, Veiga VC, et al.**
Effect of Dexamethasone on Days Alive and Ventilator-Free in Patients With Moderate or Severe Acute Respiratory Distress Syndrome and COVID-19: The CoDEX Randomized Clinical Trial. *JAMA.* 2020 Oct 6;324(13):1307.
88. **van Paassen J, Vos JS, Hoekstra EM, Neumann KMI, Boot PC, Arbous SM.**
Corticosteroid use in COVID-19 patients: a systematic review and meta-analysis on clinical outcomes. *Crit Care.* 2020 Dec;24(1):696.
89. **Gattinoni L, Busana M, Camporota L, Marini JJ, Chiumello D.**
COVID-19 and ARDS: the baby lung size matters. *Intensive Care Med.* 2021 Jan;47(1):133-4.
90. **Marini JJ, Gattinoni L.**
Management of COVID-19 Respiratory Distress. *JAMA.* 2020 Jun 9;323(22):2329.
91. **Gattinoni L, Chiumello D, Caironi P, Busana M, Romitti F, Brazzi L, et al.**
COVID-19 pneumonia: different respiratory treatments for different phenotypes? *Intensive Care Med.* 2020 Jun;46(6):1099-102.
92. **Beloncle FM, Pavlovsky B, Desprez C, Fage N, Olivier P-Y, Asfar P, et al.**
Recruitability and effect of PEEP in SARS-Cov-2-associated acute respiratory distress syndrome. *Ann Intensive Care.* 2020 Dec;10(1):55.
93. **Marini JJ, Gattinoni L.**
Management of COVID-19 Respiratory Distress. *JAMA.* 2020 Jun 9;323(22):2329-30.
94. **Abourida Y, Rebahi H, Chichou H, Fenane H, Msougar Y, Fakhri A, et al.**
What Open-Lung Biopsy Teaches Us about ARDS in COVID-19 Patients: Mechanisms, Pathology, and Therapeutic Implications. *BioMed Res Int.* 2020 Dec 10;2020:e2909673.

قسم الطبيب

أقسم بالله العظيم

أن أراقب الله في مهنتي.

وأن أصون حياة الإنسان في كافة أطوارها في كل الظروف

والأحوال باذلة وسعي في انقاذها من الهلاك والمرض

والألم والقلق.

وأن أحفظ للناس كرامتهم، وأستر عورتهم، وأكتم سرهم.

وأن أكون على الدوام من وسائل رحمة الله، باذلة رعايتي الطبية للقريب والبعيد،

للصالح والطالح، والصديق والعدو.

وأن أثابر على طلب العلم، وأسخره لنفع الإنسان لا لأذاه.

وأن أوقر من علمني، وأعلم من يصغرنني، وأكون أختاً لكل زميل في المهنة

الطبية متعاونين على البر والتقوى.

وأن تكون حياتي مصداق إيماني في سري وعلانياتي،

نقية مما يشينها تجاه الله ورسوله والمؤمنين.

والله على ما أقول شهيدا

ما تعلمنا عينة الرئة المفتوحة عن متلازمة الضائقة
التنفسية الحادة في مرضى كوفيد-19 :
الآليات، علم التشريح والآثار العلاجية.

الأطروحة

قدمت ونوقشت علانية يوم 2021/12/08
من طرف

السيدة ياسمين أبو الرضى

المزودة في 30 غشت 1996 بمراكش

لنيل شهادة الدكتوراه في الطب

الكلمات الأساسية:

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الأضرار السنخية المنتشرة

اللجنة

الرئيس

م. عبد الناصر السمكاوي

السيد

أستاذ في الإنعاش والتخدير

ح. الرايس

السيدة

أستاذة في التشريح المرضي

ح. الرباحي

السيد

أستاذ مبرز في الإنعاش والتخدير

ي. الزروقي

السيد

أستاذ مبرز في الإنعاش والتخدير

هـ. فنان

السيد

أستاذ مبرز في جراحة الصدر

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